



## TWINNER AND TENDRILLAR CLIMBER WEALTH OF VARANASI DISTRICT, UTTAR PRADESH, INDIA WITH THE ALTERNATIVE RESOURCES WITH SPECIAL EMPHASIS ON CLIMBER-BASED ANTIMICROBIALS

**Vinay Kumar Singh<sup>1</sup>, Soumyadeep Rajwar<sup>1</sup>, Raghvendra Singh<sup>1\*</sup> and Balmukund Chaurasia<sup>2</sup>**

<sup>1</sup>Centre of Advanced Study in Botany, Institute of Science, Banaras Hindu University, Varanasi - 221 005, Uttar Pradesh, India.

<sup>2</sup>H.N.754 D, Ramjankinagar, Mirzapur Pachpedwa, West Basarapatpur, Gorakhpur - 273 004, U.P., India.

\*Corresponding author E-mail : drsinghtaxon@gmail.com, singhr.bot@bhu.ac.in

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### ABSTRACT

The survey of twinner and tendrillar type of herbaceous and woody climbers was done in Varanasi district, Uttar Pradesh, India. During this survey we found twinner and tendrillar plants belong to 13 Dicotyledonous families (Acanthaceae, Apocynaceae, Aristolochiaceae, Bignoniaceae, Celastraceae, Convolvulaceae, Cucurbitaceae, Fabaceae, Menispermaceae, Passifloraceae, Polygonaceae, Ranunculaceae and Vitaceae) and 3 Monocotyledonous families (Asparagaceae, Dioscoreaceae and Smilacaceae). Most of the climbers are recorded from Convolvulaceae, Cucurbitaceae and Fabaceae. Herbaceous vines are more abundant than woody climbers (lianas). Most of the plants show twining type of climbing habit. These climbers have immense potential to combat bacterial, fungal, protozoan and viral disease without any known side effect. This present communication might be useful to researchers and policymakers to develop an alternative and effective strategy to develop drug against pathogens.

**Key words :** Angiosperm, Antimicrobial activities, Climbers, Varanasi.

### Introduction

India is one of the seventeen mega-diverse countries in the world. It is also one of the richest hotspot in the world in regard to the genetic resources of medicinal plants (Bagyalakshmi *et al.*, 2009). To available estimates, the total number of higher plant species (Angiosperms and Gymnosperms) in the world is approximately 250,000 species. Of them, only 6% have been reportedly screened for their biological activities (Katiyar *et al.*, 2012). Plants, on the basis of their growth habit, can be classified into several types e.g. herbs, shrubs, trees, climbers, etc. Climbers are plants rooted in the ground but their weak stem depends on the availability of external physical support to enhance light acquisition by preventing shading by neighboring plants (Gianoli, 2004; Suthari *et al.*, 2014). There are mainly two subdivisions of climbers: herbaceous and woody

climbers. The technical term for herbaceous climber is ‘Vine’, whereas for a woody climber ‘Liana’, which people often incorrectly used as a synonym. Climber plants also show enormous diversity in their climbing mechanisms (Putz, 1984; Bongers *et al.*, 2005). They include twiners, tendril climbers, root climbers and scramblers (Dvivedi *et al.*, 2016). A number of families such as Convolvulaceae, Cucurbitaceae and Dioscoreaceae are exclusively climbers, while more than 50 plant families like Apocynaceae, Celastraceae, Fabaceae and, Rubiaceae etc. are also have good number of climber plant species. Besides their aesthetic use, these climbers are of high medicinal value as almost all contain pharmaceutically active bio-compounds (Ali *et al.*, 2016).

Systematic investigations have been undertaken by several workers to screen the antimicrobial activities of various climber plants against numerous clinically

important infectious micro-organisms. In this present study, we surveyed and reviewed the wealth of herbaceous and woody climbers with twinning and tendrillar type of climbing patterns from Varanasi district, Uttar Pradesh, India. Their antimicrobial properties are also mentioned against highly infectious micro-organisms (Table 1), which will be extremely valuable for undertaking advance studies. The key objective of this review is to add information about twinner and tendrillar type of climber wealth of Varanasi district and to emphasize on the usage of these climbers in the selection of candidate plant species in antimicrobial research as well as to provide reference resource materials to researchers, students and policymakers in the field of drug discovery.

### Materials and Methods

Varanasi is an ancient city situated on the bank of the river Ganges, located in the southeastern part of Uttar Pradesh, India. It lies at 25.20° North latitude and 83.00° East longitude, approximately 79.1 meters above the sea level. The climate of Varanasi district is of tropical monsoonal type and experience three distinct seasons viz., cold winter (November–February), hot summer (March–June) and warm rainy seasons (June–September). The soil is of alluvial type.

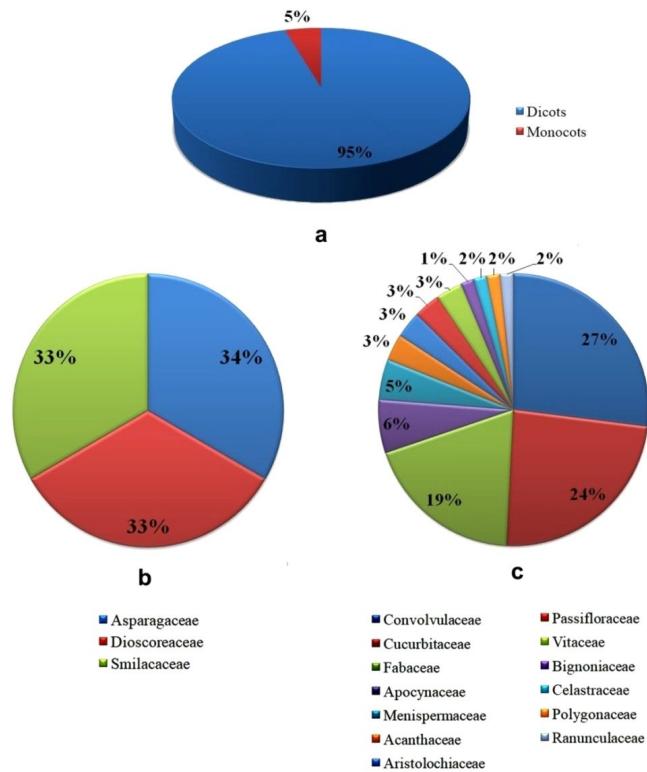
The rigorous field survey was done during the year 2018–2019 for the observation of twinner and tendrillar climber plant species of Varanasi district. A thorough survey of literature was also done for taking an account of the previously reported twinner and tendrillar plant species for Varanasi districts (Bajpai *et al.*, 1995; Dubey, 2004; Singh, 2015). Previously unreported plants species were collected and herbarium sheets were prepared using standard methods (Jain and Rao, 1978). All the taxon names were thoroughly checked for their current status using <http://plants of the world online.org/> and <http://www.theplantlist.org/>. The antimicrobial properties of these plants from various sources are summarized in this communication.

### Results and Discussion

This survey of twinner and tendrillar plant species covers about total 16 families, out of which 13 belongs to Dicotyledonous families (Acanthaceae, Apocynaceae, Aristolochiaceae, Bignoniaceae, Celastraceae, Convolvulaceae, Cucurbitaceae, Fabaceae, Menispermaceae, Passifloraceae, Polygonaceae, Ranunculaceae and Vitaceae) and from 3 belongs to Monocotyledonous families (Asparagaceae, Dioscoreaceae and Smilacaceae). There are 46 herbaceous climbers (vines) and 20 woody climbers (lianas). Maximum number of climbers present in

Convolvulaceae (17) followed by Cucurbitaceae (15) and Fabaceae (12). All the climbers show twinning pattern of climbing in the family Convolvulaceae, whereas tendrillar mode of climbing pattern was observed in Cucurbitaceae. The third largest climber plant family Fabaceae shows twinning climbing pattern except, *Vicia hirsuta* and *V. sativa*, where tendrillar nature are found. Moreover, climbers show exclusively twinning climbing pattern in the family Acanthaceae, Apocynaceae, Aristolochiaceae, Asparagaceae, Celastraceae, Dioscoreaceae, Menispermaceae, Passifloraceae and Ranunculaceae, whereas, tendrillar climbing pattern was solely found in the family Bignoniaceae, Polygonaceae, Vitaceae and Smilacaceae. Therefore, Fabaceae is the only family where both type of climbing pattern are found, while rest either has exclusively twidders or tendrillars.

This study with the findings in Fatehpur district (Agarwal, 2013) and in north-eastern district of Uttar Pradesh (Dvivedi *et al.*, 2016) concluded that maximum number of climber species with twidders and tendrillar climbing habit are from Convolvulaceae, Cucurbitaceae and Fabaceae families. This study also concluded that like Fatehpur district and north-eastern districts of Uttar Pradesh, in Varanasi (species with twidders and tendrillar climbing habit) woody climbers (lianas) are very less



**Fig. 1 :** Status of climbers. (a), Percent distribution of climbers as Monocot and Dicot (b), Percent distribution of climbers within Monocot families (c), Percent distribution of climbers within Dicot families.

**Table 1 :** Climbers from Varanasi district of Uttar Pradesh, India, with their climbing category, climbing mode and antimicrobial activities.

| S. no.              | Family           | Species  | Climbing category | Climbing mode | Antimicrobial activities with references  |
|---------------------|------------------|--|-------------------|---------------|---|
| <b>Dicotyledons</b> |                  |  |                   |               |   |
| 1.                  | Acanthaceae      | <i>Thunbergia fragrans</i> Roxb.                       | Liana             | Twinning      | Decoction of leaves shows antifungal activities (Sultana <i>et al.</i> , 2015).   |
|                     |                  | <i>Thunbergia grandiflora</i> (Roxb. ex Rottler) Roxb. | Liana             | Twinning      | Methanolic extract of flower has antibacterial activities against <i>Bacillus cereus</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Proteus mirabilis</i> , <i>Staphylococcus aureus</i> and <i>Streptococcus pyogenes</i> . Methanolic extract of leaves shows antifungal activities and anthelmintic activities against <i>Tubifex tubifex</i> (Sultana <i>et al.</i> , 2015). |
| 2.                  | Apocynaceae      | <i>Hemidesmus indicus</i> (L.) R.Br.                   | Liana             | Twinning      | Aqueous extracts from roots has significant antibacterial activity against <i>Klebsiella pneumonia</i> , <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i> (Gayathri and Kannabiran, 2009).  |
|                     |                  | <i>Ichnocarpus frutescens</i> (L.) W.T.Aiton           | Liana             | Twinning      | Chloroform root extract has significant antimicrobial activity against <i>Aspergillus flavus</i> , <i>A. niger</i> , <i>Bacillus pumilis</i> , <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> (Malathy and Sini, 2009).   |
|                     |                  | <i>Pergularia daemia</i> (Forssk.) Chiov               | Vine              | Twinning      | Methanol extract has antimicrobial activity against <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> (Doss and Anand 2013). Ethyl acetate and ethanol extracts have antibacterial activity against <i>Aeromonas hydrophila</i> , <i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> , <i>Salmonella typhi</i> and <i>Staphylococcus aureus</i> (Karthishwaran and Mirunalini, 2010).   |
|                     |                  | <i>Telosma pallida</i> (Roxb.) Craib                   | Vine              | Twinning      | Isolated alkaloids have potential antimicrobial activity (Rao and Venkatachalam, 2000).   |
| 3.                  | Aristolochiaceae | <i>Aristolochia indica</i> L.                          | Liana             | Twinning      | Extract has antimicrobial activities against <i>Aspergillus niger</i> , <i>Escherichia coli</i> and <i>Pseudomonas aeruginosa</i> (Kumar <i>et al.</i> , 2006).   |
|                     |                  | <i>Aristolochia littoralis</i> Parodi                  | Liana             | Twinning      | Extract has antimicrobial property against <i>Entamoeba histolytica</i> , <i>Giardia lamblia</i> and <i>Mycobacterium tuberculosis</i> (Jiménez-Arellanes <i>et al.</i> , 2012).  |
| 4.                  | Bignoniacae      | <i>Dolichandra unguis-cati</i> (L.) L. G.Lohmann       | Liana             | Tendrillar    | Decoction of entire plant is antimycotic for vaginal mycosis (Rondina <i>et al.</i> , 2010). Dichloromethane extract of plant is effective against <i>Staphylococcus aureus</i> (Haag <i>et al.</i> , 2014).  |
| 5.                  | Celastraceae     | <i>Celastrus paniculatus</i> Willd.                    | Liana             | Twinning      | Ethanolic extracts are effective against <i>Staphylococcus aureus</i> (Dubey <i>et al.</i> , 2012; Harish <i>et al.</i> , 2007).  |
| 6.                  | Convolvulaceae   | <i>Convolvulus arvensis</i> L.                         | Vine              | Twinning      | Extract shows significant antifungal activity against <i>Candida albicans</i> (Hassawi and Kharma, 2006). Stem and leaf extracts have antibacterial activity against <i>Escherichia coli</i> (Khan and Hayat, 2015).  |

*Table 1 continued...*

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|  |   |      |         |  |
|--|---|------|---------|--|
|  | <i>Cuscuta campestris</i> .<br>Yunck                | Vine | Twining | Lupeol epoxid isolated from this plant shows week anti HIV activity (Etedali <i>et al.</i> , 2014).  |
|  | <i>Cuscuta reflexa</i> Roxb.                        | Vine | Twining | It has antimicrobial property against the growth of <i>Staphylococcus aureus</i> (Kalita and Saikia, 2012). Methanol fraction of stem shows significant antibacterial activity against <i>Escherichia coli</i> , <i>Shigella boydii</i> , <i>S. dysenteriae</i> , <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i> (Pal <i>et al.</i> , 2006).   |
|  | <i>Ipomoea batatas</i> (L.) Lam.                    | Vine | Twining | Extract is effective in inhibition on the growth of <i>Salmonella enteritidis</i> (Cevallos-Casals and Cisneros-Zevallos, 2001).   |
|  | <i>Ipomoea cairica</i> (L.) Sweet                   | Vine | Twining | Methanol extract of leaves has significant antibacterial property against <i>Bacillus subtilis</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Salmonella typhi</i> and antifungal property against <i>Aspergillus niger</i> , <i>Candida albicans</i> , <i>Penicillium chrysogenum</i> and <i>Saccharomyces cerevisiae</i> (Srivastava and Shukla, 2015).                         |
|  | <i>Ipomoea coptica</i> (L.) Roth ex Roem. & Schult. | Vine | Twining | N/A  |
|  | <i>Ipomoea dichroa</i> Hochst. ex Choisy            | Vine | Twining | N/A  |
|  | <i>Ipomoea eriocarpa</i> R. Br.                     | Vine | Twining | Chloroform extract shows antimicrobial activity against <i>Aspergillus niger</i> and <i>Escherichia coli</i> (Das <i>et al.</i> , 2015).   |
|  | <i>Ipomoea muricata</i> (L.) Jacq.                  | Vine | Twining | Hydroalcoholic extract of the seeds has antibacterial activity against <i>Bacillus subtilis</i> , <i>Mycobacterium</i> sp., <i>Staphylococcus aureus</i> , <i>S. saprophyticus</i> and <i>Streptococcus viridians</i> (Cardona and Ysrael, 2012; Solevila and Guevara, 1991).  |
|  | <i>Ipomoea pes-tigridis</i> L.                      | Vine | Twining | The ethyl acetate extract and n-hexane extract has antimicrobial activity against <i>Bacillus subtilis</i> , <i>Staphylococcus aureus</i> and <i>Vibrio cholera</i> (Chowdhury <i>et al.</i> , 2014). Alkaloids and flavonoids of aerial parts have potential inhibitors of <i>Propionibacterium acnes</i> and <i>Staphylococcus epidermidis</i> (Sandhya <i>et al.</i> , 2012).                     |
|  | <i>Ipomoea quamoclit</i> L.                         | Vine | Twining | Ethanol extract has antimicrobial activity against gram positive bacteria ( <i>Bacillus subtilis</i> , <i>Staphylococcus aureus</i> and <i>S. warneri</i> ), gram negative bacteria ( <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Proteus mirabilis</i> , <i>Pseudomonas aeruginosa</i> and <i>P. putida</i> ) and fungal strain ( <i>Candida albicans</i> ) (Paul and Sinha, 2016). |
|  | <i>Merremia emarginata</i> (Burm.f.) Hallier f.     | Vine | Twining | Methanol extract of leaves is effective against <i>Bacillus cereus</i> and <i>Escherichia coli</i> , whereas aqueous extract is effective against <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i> (Elumalai <i>et al.</i> , 2011).  |
|  | <i>Merremia hederacea</i> (Burm.f.) Hallier f.      | Vine | Twining | N/A  |

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|                  |   |       |            |   |
|------------------|---|-------|------------|---|
|                  | <i>Operculina turpethum</i><br>(L.) Silva Manso | Liana | Twinning   | The extract has slight antimicrobial activity against <i>Staphylococcus aureus</i> (Ahmed <i>et al.</i> , 2013).  |
|                  | <i>Poranopsis paniculata</i><br>(Roxb.) Roberty | Liana | Twinning   | N/A   |
|                  | <i>Rivea hypocrateiformis</i><br>(Desr.) Choisy | Vine  | Twinning   | Crude plant extract has antimicrobial activity against <i>Bacillus cereus</i> , <i>Bacillus subtilis</i> , <i>Escherichia coli</i> and <i>Pseudomonas aeruginosa</i> (Venkata <i>et al.</i> , 2012). Aqueous extract has strong inhibitory effect against <i>Aspergillus flavus</i> , <i>A. niger</i> and <i>Candida albicans</i> (Saboo <i>et al.</i> , 2014).   |
|                  | <i>Merremia tridentata</i> (L.) Hallier f.      | Vine  | Twinning   | Methanol extracts have antimicrobial activity against <i>Aspergillus niger</i> , <i>Bacillus subtilis</i> , <i>Escherichia coli</i> , and <i>Staphylococcus aureus</i> (Kaladhar <i>et al.</i> , 2009).   |
| 7. Cucurbitaceae | <i>Benincasa hispida</i><br>(Thunb.) Cogn.      | Vine  | Tendrillar | Seed oil has antibacterial activity against <i>Bacillus subtilis</i> and methanolic extract has significant inhibition activity against <i>Candida albicans</i> (Al-Snafi, 2013).   |
|                  | <i>Bryonia cretica</i> L.                       | Vine  | Tendrillar | Root lipid extracts has antibacterial activity against <i>Enterococcus faecium</i> and <i>Streptococcus agalactiae</i> (Dhouioui <i>et al.</i> , 2016).   |
|                  | <i>Citrullus colocynthis</i> (L.) Schard.       | Vine  | Tendrillar | Ethanolic extract has inhibitory activity against <i>Staphylococcus aureus</i> (Najafi <i>et al.</i> , 2010). Antimycotic activities of the ethanol extracts are effective against <i>Alternaria alternata</i> and <i>Rhizoctonia solani</i> (Hadizadeh <i>et al.</i> , 2009).  |
|                  | <i>Coccinia grandis</i> (L.) Voigt              | Vine  | Tendrillar | Ethanolic extract has remarkable antifungal activities against <i>Aspergillus niger</i> and <i>Candida albicans</i> . Aqueous extract has remarkable antibacterial activities against <i>Bacillus subtilis</i> , <i>Escherichia coli</i> , <i>Salmonella choleraesuis</i> , <i>Shigella dysenteriae</i> and <i>S. flexneri</i> (Bhattacharya <i>et al.</i> , 2010).   |
|                  | <i>Mukia maderaspatana</i> L.                   | Vine  | Tendrillar | Chloroform and ethanol leaf extract has antimicrobial activity against <i>Aspergillus flavus</i> , <i>Klebsiella pneumoniae</i> and <i>Rhizopus</i> sp. Acetone and ethanol stem extracts have antimicrobial property against <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> and <i>Pseudomonas putida</i> (Sagayara <i>et al.</i> , 2011).   |
|                  | <i>Cucumis melo</i> L.                          | Vine  | Tendrillar | N/A   |
|                  | <i>Cucumis sativus</i> L.                       | Vine  | Tendrillar | Phosphate buffered saline (PBS) peel extract has active agents against <i>Staphylococcus aureus</i> . PBS pulp extracts has antimicrobial action against <i>Klebsiella pneumoniae</i> (Foong <i>et al.</i> , 2015).   |
|                  | <i>Cucurbita maxima</i><br>Duchense             | Vine  | Tendrillar | Ethyl acetate extract of flowers has antimicrobial activity against <i>Bacillus cereus</i> , <i>Candida albicans</i> , <i>Cochliobolus lunatus</i> , <i>Escherichia coli</i> , <i>Enterococcus faecalis</i> and <i>Salmonella typhi</i> (Muruganantham <i>et al.</i> , 2016).   |
|                  | <i>Cucurbita moschata</i><br>Duchense           | Vine  | Tendrillar | Methanolic seed extracts have antibacterial action against <i>Bacillus subtilis</i> , <i>Escherichia coli</i> <i>Klebsiella pneumoniae</i> and <i>Staphylococcus aureus</i> . Methanolic extracts of oil from seeds have antifungal activity against <i>Aspergillus parasiticus</i> , <i>Candida albicans</i> , <i>Penicillium chrysogenum</i> and <i>Rhodotorula rubra</i> (Abd El-Aziz and Abd El-Kalek, 2011). |

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|             |  |       |            |   |
|-------------|--|-------|------------|---|
|             | <i>Lagenaria siceraria</i><br>(Molina) Standl. | Vine  | Tendrillar | Phosphate buffered saline (PBS) seed extracts has antimicrobial action against <i>Escherichia coli</i> , <i>Fusarium oxysporum</i> , <i>Serratia marcescens</i> , <i>Streptococcus thermophilus</i> and <i>Trichoderma reesei</i> (Sood <i>et al.</i> , 2012).  |
|             | <i>Luffa acutangula</i> (L.) Roxb.             | Vine  | Tendrillar | Methanol fruit powder extract has antimicrobial activity against <i>Bacillus subtilis</i> , <i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> , and <i>Staphylococcus aureus</i> (Mustarchie <i>et al.</i> , 2012).   |
|             | <i>Momordica balsamina</i> L.                  | Vine  | Tendrillar | Both aqueous and alcohol extract has antimicrobial activity against <i>Bacillus subtilis</i> and <i>S. paratyphi</i> (Akinyemi, 2000). It also shows antiplasmoidal activity (Benoit-Vical <i>et al.</i> , 2006).   |
|             | <i>Momordica charantia</i> L.                  | Vine  | Tendrillar | Seed essential oil has high antimicrobial activity against <i>Staphylococcus aureus</i> and low antimicrobial activity against <i>Candida albicans</i> and <i>Escherichia coli</i> (Braca <i>et al.</i> , 2008).  |
|             | <i>Trichosanthes cucumerina</i> L.             | Vine  | Tendrillar | Cold ethanolic extract significantly inhibits the growth of <i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> , <i>Staphylococcus aureus</i> and <i>Streptococcus pyogenes</i> (Arawawala <i>et al.</i> , 2011).  |
|             | <i>Trichosanthes dioica</i> Roxb.              | Vine  | Tendrillar | Leaf, fruit and seed extract has antimicrobial activity against <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Mycobacterium smegmatis</i> , <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i> (Rai <i>et al.</i> , 2010).  |
| 8. Fabaceae | <i>Cajanus scarabaeoides</i> (L.) Thours       | Vine  | Twining    | Chloroform and ethanolic extracts have considerable antimicrobial activity against <i>Aspergillus niger</i> , <i>Bacillus subtilis</i> , <i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i> (Pattanayak <i>et al.</i> , 2009).  |
|             | <i>Canavalia gladiata</i> (Jacq.) DC.          | Vine  | Twining    | Ethanol/ methanol extracts have antimicrobial activity against <i>Klebsiella pneumonia</i> (Parekh and Chanda, 2007). Ethanol/ methanol extracts show antimicrobial activity against <i>Shigella sonnei</i> and <i>Vibrio parahemolyticus</i> (Chung <i>et al.</i> , 2014).   |
|             | <i>Clitoria ternatea</i> L.                    | Liana | Twining    | Extracts has inhibitory effects against <i>Aeromonas formicans</i> , <i>A. hydrophila</i> , <i>Bacillus subtilis</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumonia</i> , <i>Pseudomonas aeruginosa</i> , and <i>Streptococcus agalactiae</i> (Al-Snafi, 2016).  |
|             | <i>Derris scandens</i> (Roxb.) Benth.          | Liana | Twining    | Aqueous extract has antimicrobial activity against <i>Escherichia coli</i> , <i>Staphylococcus aureus</i> and <i>S. epidermidis</i> (Sittiwit and Puangprongpitag, 2009).   |
|             | <i>Lablab purpureus</i> (L.) Sweet             | Vine  | Twining    | Extracts has moderate to good antimicrobial activity against <i>Aspergillus niger</i> , <i>Bacillus megaterium</i> , <i>B. subtilis</i> , <i>Candida albicans</i> , <i>Escherichia coli</i> , <i>Saccharomyces cerevaceae</i> , <i>Salmonella paratyphi</i> , <i>S. typhi</i> , <i>Sarcina lutea</i> , <i>Shigella boydii</i> , <i>S. dysenteriae</i> , <i>Staphylococcus aureus</i> , <i>Vibrio mimicus</i> and <i>V. parahaemolyticus</i> (Al-Snafi, 2017). |

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|     |   |                                       |            |  |   |
|-----|---|---------------------------------------|------------|--|---|
|     | <i>Mucuna pruriens</i> (L.) DC.                       | Liana                                 | Twinning   | Leaf extract has strong antibacterial activity against <i>Bacillus subtilis</i> , <i>Escherichia coli</i> , <i>Proteus mirabilis</i> <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i> (Salau and Odeleye, 2007).   |   |
|     | <i>Phaseolus lunatus</i> L.                           | Vine                                  | Twinning   | N/A  |   |
|     | <i>Rhynchosia capitata</i> (B.Heyne ex Roth) DC.      | Vine                                  | Twinning   | N/A  |   |
|     | <i>Rhynchosia minima</i> (L.) DC.                     | Vine                                  | Twinning   | N/A  |   |
|     | <i>Vicia hirsuta</i> (L.) Gray                        | Vine                                  | Tendrillar | N/A  |   |
|     | <i>Vicia sativa</i> L.                                | Vine                                  | Tendrillar | n-hexane extract has antibacterial activity against <i>Bacillus atrophaeus</i> , <i>Escherichia coli</i> , <i>Staphylococcus aureus</i> , and <i>S. epidermidis</i> (Saleem et al., 2014).   |   |
|     | <i>Vigna unguiculata</i> (L.) Walp.                   | Vine                                  | Twinning   | Acetone extracts have antimicrobial activity against <i>Bacillus cereus</i> , <i>B. subtilis</i> , <i>Enterobacter cloacae</i> , <i>Enterococcus faecalis</i> and <i>Staphylococcus aureus</i> (Kritzinger et al., 2005).  |   |
| 9.  | <i>Cissampelos pareira</i> L.                         | Liana                                 | Twinning   | Methanol root extract has a broad spectrum inhibition activity for gram positive ( <i>Staphylococcus aureus</i> , <i>S. pneumoniae</i> ) and gram negative bacteria ( <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Proteus vulgaris</i> and <i>Salmonella typhimurium</i> ) (Ngoci et al., 2014).             |   |
|     | <i>Coccinia hirsutus</i> (L.) W.Theob.                | Liana                                 | Twinning   | Petroleum ether extract from dried leafs is effective against <i>Escherichia coli</i> (Satish et al., 2010). Activity against <i>Salmonella typhi</i> and <i>Staphylococcus aureus</i> were also reported to be high from dried root petroleum ether extract (Nayak and Singhai, 2003).                                      |   |
|     | <i>Tinospora cordifolia</i> (Wild.) Hook.f. & Thomson | Liana                                 | Twinning   | The ethanolic extract has significant antibacterial activity against <i>Escherichia coli</i> and <i>Proteus vulgaris</i> (Ieyachandran et al., 2003).  |   |
| 10. | <i>Passiflora edulis</i> Sims                         | Vine                                  | Twinning   | Ripe and unripe fruit peel extract has antimicrobial activity against <i>Bacillus subtilis</i> , <i>B. cereus</i> , <i>Escherichia coli</i> , <i>Lactobacillus bulgaricus</i> , <i>Proteus vulgaricus</i> , <i>Pseudomonas aeruginosa</i> , <i>Salmonelli typhi</i> and <i>Staphylococcus aureus</i> (Mohamed et al., 1994). |   |
|     | <i>Passiflora foetida</i> L.                          | Vine                                  | Twinning   | Leaf extracts have remarkable activity against <i>Pseudomonas putida</i> , <i>Shigella flexneri</i> , <i>Streptococcus pyogenes</i> , and <i>Vibrio cholera</i> (Mohanasundari et al., 2007).  |   |
| 11. | <i>Polygonum leptopus</i> Hook. & Arn.                | Vine                                  | Tendrillar | Leaf extract has significant antibacterial effect against <i>Bacillus licheniformis</i> , <i>B. subtilis</i> , <i>Streptococcus pneumonia</i> and antifungal activity against <i>Aspergillus flavus</i> , <i>A. niger</i> and <i>Candida albicans</i> (Sravanthi et al., 2017).  |   |
| 12. | <i>Ranunculaceae</i>                                  | <i>Clematis gouriana</i> Roxb. ex DC. | Liana      | Twinning   | Methanol extract has significant antimicrobial activity against <i>Candida albicans</i> , <i>Microsporum gypseum</i> , <i>M. audouinii</i> , <i>Pseudomonas aeruginosa</i> , <i>Staphylococcus aureus</i> , <i>Trichophyton rubrum</i> and <i>T. tonsurans</i> (Naika and Krishna, 2007). |

*Table 1 continued...*

**Table 1 continued...**

|                       |               |                                   |       |            |   |
|-----------------------|---------------|-----------------------------------|-------|------------|---|
| 13.                   | Vitaceae      | <i>Cayratia trifolia</i> L.       | Vine  | Tendrillar | The plant is reported to have antibacterial, antifungal and antiprotozoal activities (Kumar <i>et al.</i> , 2011).  |
|                       |               | <i>Vitis vinifera</i> L.          | Liana | Tendrillar | Aqueous and ethanolic extracts have antimicrobial activities against <i>Alcaligenes faecalis</i> , <i>Bacillus cereus</i> , <i>B. subtilis</i> , <i>Klebsiella aerogenes</i> , <i>K. pneumoniae</i> , <i>Proteus mirabilis</i> , <i>Pseudomonas pseudoalcaligenes</i> , <i>Staphylococcus aureus</i> , <i>S. epidermidis</i> and <i>S. subfava</i> (Panekh and Chanda, 2006). |
| <b>Monocotyledons</b> |               |                                   |       |            |   |
| 14.                   | Asparagaceae  | <i>Asparagus racemosus</i> Willd. | Vine  | Twining    | Methanolic extract of leaves have antimicrobial activity against <i>Bacillus subtilis</i> , <i>Candida utilis</i> , <i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i> (Patel and Patel, 2013).   |
| 15.                   | Dioscoreaceae | <i>Dioscorea bulbifera</i> L.     | Liana | Twining    | Petroleum ether and chloroform extracts has significant antimicrobial activity against <i>Aspergillus fumigatus</i> and <i>Rhizopus nigricans</i> . It has also good activity against <i>Klebsiella pneumoniae</i> . The chloroform extract has weak activity against <i>Staphylococcus aureus</i> (Seetharam <i>et al.</i> , 2003).  |
| 16.                   | Smilacaceae   | <i>Smilax zeylanica</i> L.        | Liana | Tendrillar | Leaves have antibacterial activity against <i>Escherichia coli</i> , <i>Salmonella typhi</i> , <i>S. paratyphi</i> , <i>Shigella dysenteriae</i> and <i>Staphylococcus aureus</i> (Hossain <i>et al.</i> , 2013). It was also found to be antimicrobial against <i>Aspergillus niger</i> , <i>Bacillus cereus</i> and <i>Bipolaris</i> sp. (Shree <i>et al.</i> , 2018).      |

abundant than herbaceous ones (Vines). This study extends its support to the findings of Suthari *et al.* (2014) in northern Telangana that is most of the plants in Varanasi show twinning climbing habit like northern Telangana.

Review of literatures showed that these climbers have significant antimicrobial property against numerous highly infectious bacteria (*Bacillus cereus*, *B. pumilis*, *B. subtilis*, *Enterococcus faecalis*, *Escherichia coli*, *Klebsiella pneumoniae*, *Mycobacterium tuberculosis*, *Pseudomonas aeruginosa*, *Salmonella enteritidis*, *S. typhi*, *Shigella boydii*, *S. dysenteriae*, *S. flexneri*, *S. sonnei*, *Staphylococcus aureus*, *Streptococcus pyogenes* and *Vibrio cholera* etc.), fungi (*Alternaria alternata*, *Aspergillus flavus*, *A. fumigates*, *A. niger*, *A. parasiticus*, *Candida albicans*, *C. utilis*, *Fusarium oxysporum*, *Microsporum audouinii*, *Penicillium chrysogenum*, *Rhizoctonia solani*, *Rhizopus nigrican*, *Trichoderma reesei*, *Trichophyton rubrum* and *T. tonsurans* etc.) and protozoans (*Entamoeba histolytica* and *Giardia lamblia*, etc.). Following nine climbers viz., *Cucumis melo*, *Ipomoea coptica*, *I. dichroa*, *Merremia hederacea*, *Phaseolus lunatus*, *Poranopsis paniculata*, *Rhynchosia capitata*, *R. minima* and *Vicia hirsuta* don't have any record of antimicrobial property. They may have antimicrobial property and they have not been screened so far.

From the study, it is very clear that these climbers have immense potential to combat bacterial, fungal, protozoan and viral disease without any known side effect. This present communication might be useful to researchers and policymakers to develop an alternative and effective strategy to develop drug against pathogens.

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### References

- Abd El-Aziz, A.B. and Abd El-Kalek H.H. (2011). Antimicrobial proteins and oil seeds from pumpkin (*Cucurbita moschata*). *Nature and Science*, **9(3)**, 105–119.
- Agarwal, P. (2013). Study of useful climbers of Fatehpur, Uttar Pradesh, India. *Int. J. Pharma. Life Sci.*, **4(9)**, 2957–2962.
- Ahmed, A., Howlader M.S.I., Dey S.K., Hira A., Hossain M.H. and Uddin M.M.N. (2013). Phytochemical screening and antibacterial activity of different fractions of *Operculina turpethum* root and leaf. *Amer. J. Scientific Indust. Res.*, **4(2)**, 167–172. <https://doi.org/105251/ajsir201342167172>

- Akinyemi, K.A. (2000). Antibacterial screening of five Nigerian Medicinal plants against *S. typhi* and *S. paratyphi*. *J. Nigerian Infection Control Assoc.*, **3(1)**, 30–33.
- Ali, M., Isah T. and Dipti M.A. (2016). Climber Plants: Medicinal Importance and Conservation Strategies In: Shahzad, A., Sharma S. and Siddiqui S. (eds.). *Biotechnological strategies for the conservation of medicinal and ornamental climbers*. Springer International Publishing, pp 101–138. [https://doi.org/10.1007/978-3-319-19288-8\\_4](https://doi.org/10.1007/978-3-319-19288-8_4)
- Al-Snafi, A.E. (2013). The Pharmacological importance of *Benincasa hispida*—A review. *Int. J. Pharma Sci. Res.*, **4(12)**, 165–170.
- Al-Snafi, A.E. (2016). Pharmacological importance of *Clitoria ternatea*—A review. *IOSR J. Pharmacy*, **6(3)**, 68–83.
- Al-Snafi, A.E. (2017). The pharmacology and medical importance of *Dolichos lablab* (*Lablab purpureus*) —A review. *IOSR J. Pharmacy*, **7(2)**, 22–30. <https://doi.org/10.9790/3013-0702012230>
- Arawwawala, L.D.A.M., Thabrew I., Arambewela L.S.R., Fernando N. and Guruge L.D. (2011). Antibacterial activity of *Trichosanthes cucumerina* Linn. extracts. *Int. J. Pharmaceut. Biol. Arch.*, **2**, 808–812.
- Bagyalakshmi, B., Sridhar D. and Ponmurgan P. (2009). Antimicrobial activity of important Indian medicinal plants against pyogenic infection. *J. Phytotherapy*, **1(6)**, 391–396.
- Bajpai, A., Ojha J.K. and Sant H.R. (1995). Medicobotany of the Varanasi District, Uttar Pradesh, India. *Int. J. Pharmacog.*, **33(2)**, 172–176. <https://doi.org/10.3109/13880209509055220>
- Benoit-Vical, F., Grellier P., Abdoulaye A., Moussa I., Ousmane A., Berry A., Ikhiri K. and Poupat C. (2006). *In vitro* and *in vivo* antiplasmodial activity of *Momordica balsamina* alone or in a traditional mixture. *Cancer Chemotherapy*, **52(6)**, 288–292. <https://doi.org/10.1159/000095960>
- Bhattacharya, B., Samanta M., Pal P., Chakraborty S. and Samanta A. (2010). Evaluation of Antifungal and Antibacterial Activities of the Plant *Coccinia grandis* (L.) Voigt. (Family-Cucurbitaceae). *J. Phytotherapy* **2(11)**, 52–57.
- Bongers, F., Parren M.P. and Traor D. (Eds.) (2005). Forest climbing plants of West Africa: diversity ecology and management. CAB International Wallingford Oxfordshire UK, pp 5–18. <https://doi.org/10.1079/97808519991420000>
- Braca, A., Siciliano T.D., Arrigo M. and Germanò M.P. (2008). Chemical composition and antimicrobial activity of *Momordica charantia* seed essential oil. *Fitoterapia*, **79(2)**, 123–125. <https://doi.org/10.1016/j.fitote.2007.11.002>
- Cardona, C.C. and Ysrael M.C. (2012). Antimicrobial activity of the crude ethanol extract of the seeds of *Ipomoea muricata* (Jacq.), Convolvulaceae against selected clinical isolates. *Acta Manilana*, **60**, 49–55.
- Cevallos-Casals, B.A. and Cisneros-Zevallos L.A. (2001). Bioactive and functional properties of purple sweetpotato [*Ipomoea batatas* (L.) Lam.]. *International Conference on Sweet potato Food and Health for the Future*, **583**, 195–203. <https://doi.org/10.17660/ActaHortic200258322>
- Chowdhury, R.H., Saha R., Bhuiyan M.I., Hossain M.A., Kowsar S.A.M. and Hossain M.M. (2014). An *In vitro* Assessment of Antimicrobial Thrombolytic and Cytotoxic Activity on *Ipomoea pestigridis*. *J. Advancement Medical Life Sci.*, **2(1)**, 1–8.
- Chung, J., Lee J. and Ha D. (2014). Antimicrobial activities of sword bean (*Canavalia gladiata*) extracts against food poisoning bacteria. *J. Food Hyg. Saf.*, **29(4)**, 376–382. <https://doi.org/10.13103/JFHS2014294376>
- Das, M., Babu G.S., Vidya R. and Himaja M. (2015). GC-MS analysis, antimicrobial and insecticidal activity of the leaves of *Ipomoea eriocarpa*. *J. Indian Chem. Soc.*, **92**, 542–544.
- Dhouioui, M., Boulila A., Jemli M., Schiets F., Casabianca H. and Zina M.S. (2016). Fatty acids composition and antibacterial activity of *Aristolochia longa* L. and *Bryonia dioica* Jacq. growing wild in Tunisia. *J. Oleo Science*, **65(8)**, 655–661. <https://doi.org/10.5650/josess16001>
- Doss, A. and Anand S.P. (2013). Antimicrobial activity of *Hygrophila auriculata* (Schumach.) Heine and *Pergularia daemia* Linn. *Africa J. Plant Sci.*, **7(4)**, 137–142. <https://doi.org/10.5897/AJPS12193>
- Dubey, D., Sahu M.C., Rath S., Paty B.P., Debata N.K. and Padhy R.N. (2012). Antimicrobial activity of medicinal plants used by aborigines of Kalahandi, Orissa, India against multidrug resistant bacteria. *Asian Pac. J. Trop. Biomed.*, **2(2)**, S846–S854. [https://doi.org/10.1016/S2221-1691\(12\)60322-0](https://doi.org/10.1016/S2221-1691(12)60322-0)
- Dubey, N.K. (2004). *Flora of BHU campus*. Banaras Hindu University.
- Dvivedi, A., Srivastava S. and Shukla R.P. (2016). Climber Diversity across Vegetational Landscape of North-Eastern Uttar Pradesh, India. *Notulae Scientia Biologicae*, **8(4)**, 489–497. <https://doi.org/10.15835/nsb849921>
- Elumalai, E.K., Ramachandran M., Thirumalai T. and Vinothkumar P. (2011). Antibacterial activity of various leaf extracts of *Merremia emarginata*. *Asian Pac. J. Trop. Biomed.*, **1(5)**, 406–408. [https://doi.org/10.1016/S2221-1691\(11\)60089-0](https://doi.org/10.1016/S2221-1691(11)60089-0)
- Etedali, P., Behbahani M., Rahiminejad R.M. and Rad S.J. (2014). Effect of crude extracts and fractions of *Cuscuta campestris* and Twinningo different hosts on peripheral blood mononuclear cells and HIV replication. *Int. J. Biosci.*, **4(9)**, 83–89. <https://doi.org/10.12692/ijb/4983-89>
- Foong, F.H.N., Mohammad A. and Ichwan S.J.A. (2015). Biological properties of cucumber (*Cucumis sativus* L.) extracts. *Malaysian J. Anal. Sci.*, **19(6)**, 1218–1222.
- Gayathri, M. and Kannabiran K. (2009). Antimicrobial activity of *Hemidesmus indicus*, *Ficus bengalensis* and *Pterocarpus marsupium*. *Indian J. Pharmaceut. Sci.*, **71(5)**, 578–581. <https://doi.org/10.4103/0250-474X58182>
- Gianoli, E. (2004). Evolution of a climbing habit promotes diversification in flowering plants. *Proc. Royal Soc.*

- London Ser. B: Biol. Sci.*, **271(1552)**, 2011–2015. <https://doi.org/10.1098/rspb.2004.2827>
- Haag, G.O., Valle M.E.D., Debenedetti S.L., Marín G., Brignoles P. and Magariños M.D.C. (2014). Antimicrobial activity of Latin American medicinal plant extracts. *J. Sci.*, **4(2)**, 128–131.
- Hadizadeh, I., Peivastegani B. and Kolahi M. (2009). Antifungal activity of nettle (*Urtica dioica* L.) colocynth (*Citrullus colocynthis* (L.) Schrad.) oleander (*Nerium oleander* L.) and konar (*Ziziphus spina-christi* L.) extracts on plants pathogenic fungi. *Pak. J. Biol. Sci.*, **12(1)**, 58–63. <https://doi.org/10.3923/pjbs.20095863>
- Harish, B.G., Krishna V., Sharath R., Kumara S., Raja N. and Mahadevan K.M. (2007). Antibacterial activity of celapanin a sesquiterpene isolated from the leaves of *Celastrus paniculatus* Willd. *Int. J. Biomed. Pharmaceut. Sci.*, **1(1)**, 65–68.
- Hassawi, D. and Kharma A. (2006). Antimicrobial activity of some medicinal plants against *Candida albicans*. *Int. J. Biol. Sci.*, **6(1)**, 109–114. <https://doi.org/10.3923/jbs.2006109114>
- Hossain, A.M., Saha S., Asadujjaman M. and Kahan A.S. (2013). Analgesic antioxidant and antibacterial activity of *Smilax zeylanica* Linn. (family-Smilacaceae). *Pharmacologyonline*, **1**, 244–250.
- Jain, S.K. and Rao R.R. (1978). *A handbook of field and herbarium methods*. Today and Tomorrow's Publication, New Delhi.
- Jeyachandran, R., Xavier T.F. and Anand S.P. (2003). Antibacterial activity of stem extracts of *Tinospora cordifolia* (Willd.) Hook. F. & Thomson. *Ancient Science of Life*, **23(1)**, 40.
- Jiménez-Arellanes, A., León-Díaz R., Meckes M., Tapia A., Molina-Salinas G.M., Luna-Herrera J. and Yépez-Mulia L. (2012). Antiprotozoal and antimycobacterial activities of pure compounds from *Aristolochia elegans* rhizomes. *Evidence-Based Complementary and Alternative Medicine*, pp 7. <https://doi.org/10.1155/2012/593403>
- Kaladhar, D.S.V.G.K., Harasreeramulu S., Rachel K.V. and Surekha C.H. (2009). Evaluation of Antimicrobial activity of Methanolic leaf extracts of selected *in vitro* and *in vivo* grown Convolvulaceae members. *J. Pure Appl. Microbiol.*, **3(2)**:759–767
- Kalita, D. and Saikia J. (2012). Ethnomedicinal antibacterial and antifungal potentiality of *Centella asiatica*, *Nerium indicum* and *Cuscuta reflexa*—widely used in Tiwa Tribe of Morigaon district of Assam, India. *Int. J. Phytomed.*, **4**, 380–385.
- Karthishwaran, K. and Mirunalini S. (2010). Therapeutic potential of *Pergularia daemia* (Forsk.): the Ayurvedic wonder. *Int. J. Pharmacol.*, **6(6)**, 836–843. <https://doi.org/10.3923/ijp.2010836843>
- Katiyar, C., Gupta A., Kanjilal S. and Katiyar S. (2012). Drug discovery from plant sources: An integrated approach. *Ayu*, **33(1)**, 10–19. <https://doi.org/10.4103/0974-8520100295>
- Khan, M.U. and Hayat M.Q. (2015). Phytochemical analyses for antibacterial activity and therapeutic compounds of *Convolvulus arvensis* L. collected from the salt range of Pakistan. *Adv. Life Sci.*, **2(2)**, 83–90.
- Kritzinger, Q., Lall N., Aveling T.A.S. and van Wyk B.E. (2005). Antimicrobial activity of cowpea (*Vigna unguiculata*) leaf extracts. *South Afr. J. Bot.*, **71(1)**, 45–48. [https://doi.org/10.1016/S0254-6299\(15\)30147-2](https://doi.org/10.1016/S0254-6299(15)30147-2)
- Kumar, D., Kumar S., Gupta J., Arya R. and Gupta A. (2011). A review on chemical and biological properties of *Cayratia trifolia* Linn. (Vitaceae). *Pharmacog. Rev.*, **5(10)**, 184–188. <https://doi.org/10.4103/0973-784791117>
- Kumar, V.P., Chauhan N.S., Padh H. and Rajani M. (2006). Search for antibacterial and antifungal agents from selected Indian medicinal plants. *J. Ethnopharmacol.*, **107(2)**, 182–188. <https://doi.org/10.1016/j.jep.2006.03.013>
- Malathy, N.S. and Sini S. (2009). Antimicrobial activities of *Ichnocarpus frutescens* (L.) R.Br. and *Hemidesmus indicus* R.Br. Roots. *Ancient Science of Life*, **28(4)**, 13–15.
- Mohamed, S., Hassan Z. and Hamid N.A. (1994). Antimicrobial activity of some tropical fruit wastes (guava, star fruit, banana, papaya, passion fruit, langsat, duku, rambutan and rambai). *Pertanika*, **17**, 219–219.
- Mohanasingh, C., Natarajan D., Srinivasan K., Umamaheswari S. and Ramachandran A. (2007). Antibacterial properties of *Passiflora foetida* L.—a common exotic medicinal plant. *Afr. J. Biotech.*, **6(23)**, 2650–2653. <https://doi.org/10.5897/AJB2007000-2426>
- Muruganantham, N., Solomon S. and Senthamilselvi M.M. (2016). Antimicrobial activity of *Cucurbita maxima* flowers (Pumpkin). *J. Pharmacog. Phytochem.*, **5(1)**, 15–18.
- Mustarichie, R., Udin L.Z. and Muchtaridi S. (2012). Identification and antibacterial activity of methanol extract of *Luffa Acutangula* Roxb. *Int. J. Med. Hlth Sci.*, **12**, 64–71. <https://doi.org/10.15208/mhsj201249>
- Naika, H.R. and Krishna V. (2007). Antimicrobial Activity of Extracts from the Leaves of *Clematis gouriana* Roxb. *Int. J. Biomed. Pharmaceut. Sci.*, **1(1)**, 69–72.
- Najafi, S., Sanadgol N., Nejad B.S., Beiragi M.A. and Sanadgol E. (2010). Phytochemical screening and antibacterial activity of *Citrullus colocynthis* (Linn.) Schrad. against *Staphylococcus aureus*. *J. Medicinal Plants Res.*, **4(22)**, 2321–2325.
- Nayak, S. and Singhai A.K. (2003). Antimicrobial activity of the roots of *Cocculus hirsutus*. *Ancient Science of Life*, **22(3)**, 101–105.
- Ngoci, N.S., Ramadhan M., Ngari M.S. and Leonard O.P. (2014). Screening for antimicrobial activity of *Cissampelos pareira* L. methanol root extract. *Europ. J. Medicinal Plants*, **4(1)**, 45–51. <https://doi.org/10.9734/EJMP/2014/5464>
- Pal, D.K., Mandal M., Senthilkumar G.P. and Padhiari A. (2006). Antibacterial activity of *Cuscuta reflexa* stem and *Corchorus olitorius* seed. *Fitoterapia*, **77(7–8)**, 589–

591. <https://doi.org/101016/j.fitote200606015>
- Parekh, J. and Chanda S. (2006). *In-vitro* antimicrobial activities of extracts of *Launaea procumbens* Roxb. (Labiateae), *Vitis vinifera* L. (Vitaceae) and *Cyperus rotundus* L. (Cyperaceae). *Afr. J. Biomed. Res.*, **9**, 89–93. <https://doi.org/104314/ajbrv9i248780>
- Parekh, J. and Chanda S. (2007). *In vitro* screening of antibacterial activity of aqueous and alcoholic extracts of various Indian plant species against selected pathogens from *Enterobacteriaceae*. *Afr. J. Microbiol. Res.*, **1(6)**, 92–99.
- Patel, L.S. and Patel R.S. (2013). Antimicrobial activity of *Asparagus racemosus* Willd. from leaf extracts—a medicinal plant. *Int. J. Scientific Res. Publications*, **3(3)**:2250–3153
- Pattanayak, S., Sankar N.S. and Panda D.P. (2009). Antimicrobial activity of petroleum ether chloroform and ethanolic extracts of *Cajanus scarabaeoides* (L.) whole plant. *Pharmacologyonline*, **1**, 530–536.
- Paul, D. and Sinha S.N. (2016). An update on biological activities of medicinal plant *Ipomoea quamoclit* L. *Tropical Plant Res.*, **3(1)**, 186–190.
- Putz, F.E. (1984). The natural history of lianas on Barro Colorado Island Panama. *Ecology*, **65**, 1713–1724. <https://doi.org/102307/1937767>
- Rai, P.K., Mehta S., Gupta R.K. and Watal G. (2010). A novel antimicrobial agents *Trichosanthes dioica*. *Int. J. Pharma Bio Sci.*, **1(3)**, 202–209.
- Rao, K.N. and Venkatachalam S.R. (2000). Inhibition of dihydrofolate reductase and cell growth activity by the phenanthroindolizidine alkaloids pergularinine and tylophorinidine: the *in vitro* cytotoxicity of these plant alkaloids and their potential as antimicrobial and anticancer agents. *Toxicology in vitro*, **14(1)**, 53–59. [https://doi.org/101016/S0887-2333\(99\)00092-2](https://doi.org/101016/S0887-2333(99)00092-2)
- Rondina, R.V., Bandoni A.L. and Coussio J.D. (2010). Argentine medicinal plants with potential antifungal activity. *Dominguezia*, **26(1)**, 31–39.
- Saboo, S., Tapadiya G.G. and Khadabadi S.S. (2014). Antimicrobial and Phytochemical Analysis of *Revia hypocrateriformis*. *Microbiol. J.*, **4(1)**, 22–24. <https://doi.org/103923/mj20142226>
- Sagayaraj, T., Kannan N.R., Churchill M.B., Label L.A. and Natarajan E. (2011). Antimicrobial properties of *Mukia maderaspatana* L. *Indian J. Appl. Pure Biol.*, **26(1)**, 1–4.
- Salau, A.O. and Odeleye O.M. (2007). Antimicrobial activity of *Mucuna pruriens* on selected bacteria. *Afr. J. Biotech.*, **6(18)**, 2091–2092. <https://doi.org/105897/AJB2007000-2324>
- Saleem, M., Karim M., Qadir M.I., Ahmed B., Rafiq M. and Ahmad B. (2014). *In vitro* antibacterial activity and phytochemical analysis of hexane extract of *Vicia sativa*. *Bangladesh J. Pharmacol.*, **9(2)**, 189–193. <https://doi.org/103329/bjp9i217859>
- Sandhya, S., Sravanthi E.V., Vinod K.R., Gouthami G., Saikiran M. and Banji D. (2012). Alkaloids and Flavonoids of Aerial Parts of *Ipomea pes-tigridis* (Convolvulaceae) are Potential Inhibitors of *Staphylococcus epidermidis* and *Propionibacterium acnes*. *J. Herbs Spices & Medicinal Plants*, **18(4)**, 370–386. <https://doi.org/101080/104964752012715118>
- Satish, V., Ravichandran V.D., Gavani U. and Paarak P.M. (2010). Antimicrobial studies on the extracts of *Cocculus hirsutus* Linn. and *Hyptis suaveolens* Poit. *Indian J. Nat. Products and Resources*, **1**, 49–52.
- Seetharam, Y.N., Jyothishwaran G., Sujeeeth H., Barad A., Sharanabasappa G. and Shivkumar D. (2003). Antimicrobial activity of *Dioscorea bulbifera* bulbs. *Indian J. Pharmaceut. Sci.*, **65(2)**, 195–196.
- Shree, V.D., Arbin A., Noorain G.S., Sahana B.K. and Kekuda T.P. (2018). Preliminary phytochemical analysis antimicrobial and antioxidant activity of *Smilax zeylanica* L. (Smilacaceae). *J. Drug Del. Therap.*, **8(4)**, 237–243. <https://doi.org/1022270/jddtv8i41779>
- Singh, A. (2015). Observations on the flora of Varanasi district in Uttar Pradesh state of India. *Glob. J. Environ. Sci. Tech.*, **3(10)**, 368–389.
- Sittiwit, C. and Puangprongpitak D. (2009). Antimicrobial properties of *Derris scandens* aqueous extract. *J. Biol. Sci.*, **9(6)**, 607–611. <https://doi.org/103923/jbs2009607611>
- Solevilla, R.C. and Guevara B.Q. (1991). Biological studies of the seed of *Ipomoea muricata* Jacq.; Linne. (Convolvulaceae). *Substances naturelles d'origine vegetable*, 143–145
- Sood, A., Kaur P. and Gupta R. (2012). Phytochemical screening and antimicrobial assay of various seeds extract of Cucurbitaceae family. *Int. J. Appl. Biol. Pharmaceut. Tech.*, **3(3)**, 401–409.
- Sravanthi, M., Padmaja M., Muni K.D. and Hemalatha K.P.J. (2017). *In vitro* antimicrobial properties and phytochemical screening of crude extracts of *Antigonon leptopus* Hook. & Arn. leaf. *Int. J. Innov. Pharmaceut. Sci. Res.*, **5**, 1–13.
- Srivastava, D. and Shukla K. (2015). *Ipomoea cairica*: a medicinal weed with promising health benefits. *Int. J. Information Res. Rev.*, **2(5)**, 687–694.
- Sultana, K.W., Chatterjee S. and Roy C.I. (2015). An overview on Ethnopharmacological and Phytochemical properties of *Thunbergia* sp. *Medicinal and Aromatic Plants Res. J.*, **4 (5)**, 217. <https://doi.org/104172/2167-04121000217>
- Suthari, S., Sreeramulu N., Omkar K. and Raju V.S. (2014). The climbing plants of northern Telangana in India and their ethnomedicinal and economic uses. *Indian J. Plant Sci.*, **3(1)**, 86–100.
- Venkata, S.P., Murali M.C., da Silva J.A.T., Raju B.A. and Sravani R. (2012). Screening the Antimicrobial and Antioxidant Potential of *Ventilago dentate*, *Scolopia crenata*, and *Rivea hypocrateriformis* from Maredumilli Forest, India. *Medicinal and Aromatic Plant Science and Biotechnology*, **6(1)**, 58–62.