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EXOGENOUS APPLICATION OF IBA AND NAA IMPROVED ROOTING AND SURVIVAL OF HARDWOOD CUTTINGS OF FIG (*FICUS CARICA* L.) CV. DINKAR

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ABSTRACT

Fig (*Ficus carica* L.) stem-cutting propagation method is slow and a lower percentage of the cuttings finally survive due to poor rooting, making it challenging to adopt. This study investigated six distinct concentrations of Indole-3-butyric acid (IBA) and 1-naphthaleneacetic acid (NAA) with control. Treatment T₆ demonstrated the most favourable outcome, displaying a rooting percentage of 69.22%, a maximum root length of 23.18 cm, the highest number of roots of 26 per cutting, roots weights of 2.53 g (fresh) and 0.79 g (dry). Additionally, it exhibited a higher survival rate of 87.35% at 90 days after planting (DAP). Treating fig hardwood cuttings with 2000 ppm of IBA and 2000 ppm NAA and growing them under shade house conditions can be recommended to researchers and the nursery industry to significantly improve rooting success and the overall survival rate of fig cuttings.

Keywords : *Ficus carica* L., IBA, NAA, rooting, hardwood cutting

Introduction

Fig (*Ficus carica* L.) is the oldest domesticated fruit by humans belonging to the Moraceae family. It is native to the Middle East and widely distributed worldwide, including regions such as the Indian subcontinent, Mediterranean countries, South California and the Far East (Lansky & Paavilainen, 2010). Fig tree produces anjeer, a delicious fruit renowned for its exceptional nutritional qualities, consumed both fresh and dried (Kumar *et al.*, 1998). Besides, barks, leaves and fruits of this species play an important role as a medicinal tree to cure diabetes, skin diseases, ulcers, dysentery and diarrhea (Vora *et al.*, 2017). As a traditional tree, this species readily adapts to poor conditions, showing resistance to high calcium, salt, and drought soil levels.

The fig produces non-viable seeds, so these plants are generally multiplied by air-layering, cutting, grafting and tissue culture (Kumar *et al.*, 1998). Tissue culture has prerequisites, grafting takes a long time and air-layering produces fewer seedlings due to the delicate nature of the mother plant (Koyuncu *et al.*, 2003; Shamsuddin *et al.*, 2021). Cutting propagation, particularly with hardwood cuttings, can produce substantial quantities of true-to-type planting material while preventing deterioration of the mother plant. Apart from that, the stem-cutting propagation method takes a lot of time and only 20–30% of the cuttings survive due to poor rooting, making it challenging to adopt (Darwesh *et al.*, 2014).

Plant multiplication by hardwood cuttings is highly reliant on adventitious root formation (Husen *et*

al., 2017). The initiation of adventitious roots is a complicated developmental process influenced by plant growth hormones, water relations of the cuttings and their nutritional level (Hartmann *et al.*, 2017). Plant growth regulatory 'auxins' play a vital role in influencing the rooting and survival of stem cuttings. Previous reports suggested that exogenously applied auxins may enhance adventitious rooting (AR) triggering the action of endogenous auxin (Park *et al.*, 2011). Additionally, it enhances the initiation and development of root primordium through cell division (Fogaca *et al.*, 2005). Indole Butyric Acid (IBA) and Naphthalene Acetic Acid (NAA) are synthetic auxins that are reliable in the promotion of rooting in cuttings (Tsipouridis *et al.*, 2003). Furthermore, IBA is probably the ideal substance for general application since it is non-harmful to plants with a large range of concentrations and highly impactful in accelerating root formation in many plant species (Hartmann *et al.*, 2017). NAA could ensure a greater rooting capacity, cuttings' survival and reduced time required for rooting (Kaewchangwat *et al.*, 2020). These factors have an immense effect on desirable rooting and survival of propagated cuttings. Research on the improvement of propagation techniques for fig trees is of paramount importance for the development of well-being, fruitful and profitable fig orchards (Czaja *et al.*, 2016). This work aimed to determine the best combination of IBA and NAA for achieving desirable rooting with sustainable survival in fig hardwood cuttings.

Material and Methods

The present investigation was conducted at the Research Farm of Banda University of Agriculture and Technology, Banda located at 25 °31' N latitude, 80 °20' E longitude and an elevation of 146 m above sea level during the period of 2020-2021. The hardwood cuttings were obtained from cv. Dinakar (Fig. 1) as mother trees that were 2 years old, were maintained under optimal horticultural practices. The experimental research was organized using Randomized Block Design (RBD), where seven unique concentrations of IBA and NAA *i.e.*, T₁- IBA 1000 ppm, T₂- IBA 2000 ppm, T₃-NAA 1000 ppm, T₄ -NAA 2000 ppm, T₅-IBA 1000 ppm + NAA 1000 ppm, T₆- IBA 2000 ppm + NAA 2000 ppm and T₇-control were investigated. Each combination was replicated three times, and each replication consisted of 30 individual cuttings cultivated under a shade house environment (Fig.3) for the duration of the experiment. The various hormonal formulations were prepared (Fig.2.) by dissolving the required quantity of individual hormones in 10 mL of methanol and then making up the volume of 1 liter by adding distilled water. For fig plant cuttings, the basal

sections were rapidly immersed in specific solutions utilizing the quick-dip technique. During this procedure, approximately 3 cm of the cut end of each cutting were submersed in the hormonal solution for duration of 1 minute. Subsequently, the cuttings were subjected to a 15-minute period of shade drying and then planted into polybags measuring 6 cm × 9 cm in size. These polybags were filled with a mixture of soil, farmyard manure (FYM), and sand in a volumetric ratio of 1:3 (soil: FYM and sand). The polybags were kept in a shade house and irrigated on a regular basis. The parameters took into account the percentage of rooted cuttings, length of longest root (cm), number of roots (Fig. 4), fresh and dry weight (g) of roots and survival percentage were taken at 90 DAP. Recorded data of the above-mentioned parameters were subjected to statistical analysis by using SAS 2.0 (statistical analysis system software version).

Result and Discussion

Rooting percentage

The rooting percentage of cuttings was studied on the 90th day and it was observed that among the different concentrations of growth hormone, cutting treated with T₆ recorded the maximum rooted cuttings percentage (69.22%), followed by T₅ recorded 67.78% rooted cuttings as represented in Fig. 5. Moreover, the minimum rooting percentage (36.67%) observed with T₇- control. Yusnita *et al.* (2017) observed that 100% of the roots formed when IBA and NAA were combined in semi-hardwood cuttings of Malay apples. Further, In *Cordyline terminalis* and *Azalea alexander* L. higher rooting was recorded when IBA and NAA were applied in combined form rather than alone (Rahdari *et al.*, 2014; Mohana *et al.*, 2014). IBA promotes root development in stem cuttings through processes like conversion to IAA, elevating free-IBA levels, improving tissue responsiveness to IAA, boosting internal IAA synthesis, and potentially collaborating with IAA's function.

Number of roots

The significant influence of growth hormone on the number of roots in fig cuttings on the 90th day of treatment is presented in Table 1. Among various treatments, the maximum root count (26.67) was achieved with T₆, followed by T₅ with 25.13 roots and T₂ of 23.47. Conversely, the lowest root count (17.93) was observed in T₇-control. Khalid and Ahmed (2022) revealed that the rooting number increased significantly, reaching 59.85 with the application of NAA. Additionally, Reddy *et al.* (2008) recorded that the maximum number of roots were obtained when both IBA and NAA were used in combination. In

particular, the observed phenomenon might be because the use of auxins stimulates cambial activity, leading to the movement of stored food materials towards the location of root initiations.

Length of the longest root

The length of the longest root examined on 90 DAP is displayed in Table 1. Treatment T₆ recorded the longest root (23.18 cm) followed by T₅ with a measurement of 21.54 cm. Mewar and Naithani (2016) recorded that treatment with 6000 ppm of IBA in wild figs exhibited a markedly longer root length (12.13 cm). Babaie *et al.* (2014), observed the maximum root length in *Ficus binnendijkii* cuttings treated at 6000 ppm IBA. Reddy *et al.* (2008) recorded that fig hardwood cuttings treated with IBA 2500 + NAA 2500 ppm produced the longest root length per cutting (19.05 cm). Particularly, the action of IBA on the translocation of metabolites and carbohydrate metabolism, which may be implicated in the influence of hormones on root length, maybe the cause of this rise in root length. Another possibility is that auxin activity triggered the hydrolysis and transfer of carbohydrates and nitrogenous compounds located close to the base of cuttings, which speed up cell elongation and cell division in a beneficial environment (Mewar & Naithani, 2016).

Fresh and dry root weights

Ninety days after planting the treatment revealed an impressive impact of growth hormone on both the fresh and dry root weights as indicated by the graphical illustration in Fig. 6. The maximum fresh weight of roots (2.53 g) was observed in T₆, followed by T₅ (2.37 g). In contrast, the minimum fresh weight (1.64 g) was recorded in T₇.control which was statistically similar to T₃ (1.62 g). Besides the dry weight with T₆ exhibited the maximum (0.79 g), succeeded by T₅ (0.74 g) and T₂ (0.67 g). Conversely, the lowest dry weight (0.51 g) was observed in the T₇.control which is almost similar to T₃ (0.52 g) and it was followed by T₄ (0.55 g) and T₁ (0.64 g). According to several studies, the combination of IBA and NAA produces a maximum fresh and dry mass of roots in hardwood cuttings of Phalsa (Singh & Tomar, 2015; Ghosh *et al.*, 2017). The rise in root count per cutting possibly impacted root fresh weight, as evidenced in Phalsa (Singh & Tomar, 2015). Additionally, the increased dry root weight could be linked to greater root length, facilitating the accumulation of stored carbohydrates and a higher root count leading to increased root volume per cutting in hardwood.

Survival percentage

As indicated in Fig.7 there was a notable variance in survival percentage among different levels of growth hormone. Treatment T₆ was statistically on par with T₅ and T₂, and had the highest survival rate while T₇.control showed the lowest survival percentage, which was statistically comparable to T₃, followed by T₁. Moreover, the survival rate in T₆ increased by 1.15 times higher than the T₇.control. Treatment of IBA at 3000 ppm boasts considerably the highest survival rate of rooted cuttings of fig cv. Dinkar (Kuntagol *et al.*, 2018). Patel *et al.* (2017) found that the IBA @4000 ppm measured the highest survival rate (67.2 %). Besides, it could be because the roots were longer and more numerous under this treatment, which allowed for greater absorption of nutrients and moisture from the media and as a result of higher survival rate (Reddy *et al.*, 2008).

Conclusion

Based on the findings of the present study, the treatment of fig hardwood cuttings with IBA at 2000 ppm and NAA at 2000 ppm under shade house conditions is recommended as an effective and practical approach for nursery growers and researchers aiming to enhance the rooting and survival of fig hardwood cuttings. This research contributes to the optimization of propagation techniques in fig cultivation, potentially leading to increased productivity and success in fig nurseries



Fig. 1: Preparation of Hardwood cuttings of fig cv. Dinkar.



Fig. 2: Prepared solution of different concentrations of IBA and NAA.

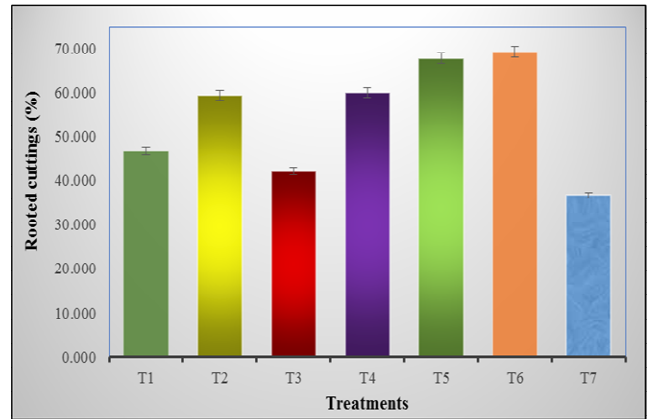


Fig. 5: Percentage of rooted cuttings in hardwood cuttings of fig (*Ficus carica* L.) cv. Dinkar at different treatments.



Fig. 3: Different treatments well managed under shade house.

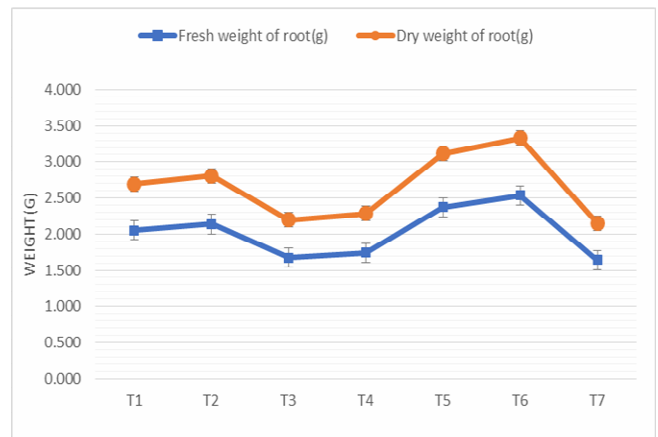


Fig. 6: Changes of the fresh and dry weight of roots of hardwood cuttings of fig (*Ficus carica* L.) cv. Dinkar at different treatments.

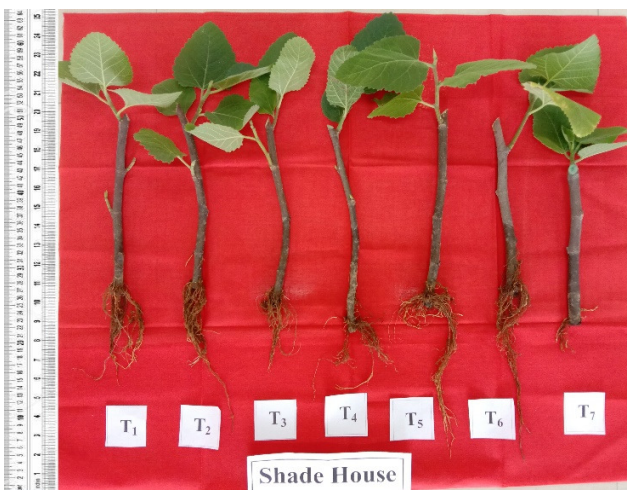


Fig. 4 : Rooting performances of hardwood cuttings of fig cv. Dinkar with different treatments.

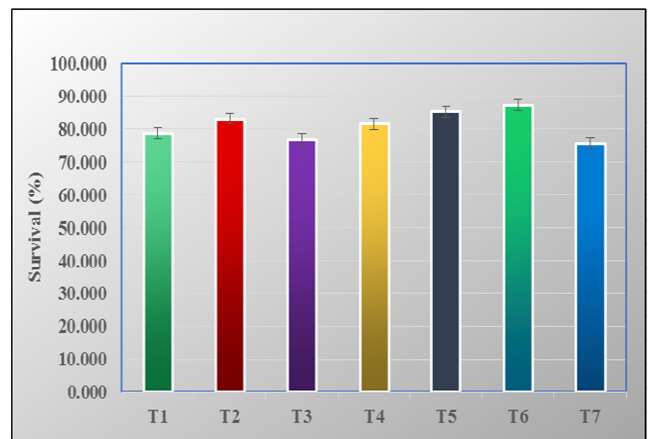


Fig. 7 : Survival trends in hardwood cuttings of fig (*Ficus carica* L.) cv. Dinkar at different treatments.

Table 1: Effect of different treatments (T₁-T₇) on the percentage of rooted cuttings, number of roots and length of longest root in hardwood cuttings of fig

Treatments	Number of roots/ cutting	Length of longest root (cm)
T ₁ (IBA @ 1000 ppm)	22.667 ^{Cc}	18.003 ^{Bbcd}
T ₂ (IBA @ 2000 ppm)	23.467 ^{Cc}	18.603 ^{Bbc}
T ₃ (NAA @ 1000 ppm)	18.933 ^{Dd}	13.940 ^{Cd}
T ₄ (NAA @ 2000 ppm)	19.533 ^{Dd}	14.730 ^{Ccd}
T ₅ (IBA @ 1000 ppm + NAA @ 1000 ppm)	25.133 ^{Bb}	21.537 ^{Aab}
T ₆ (IBA @ 2000 ppm + NAA @ 2000 ppm)	26.667 ^{Aa}	23.183 ^{Aa}
T ₇ -(Control)	17.933 ^{Ec}	14.357 ^{Ccd}
SEm±	0.283	1.486
C.D. (P=0.05)	0.872	4.579

References

- Babalie, H., Zarei, H., Nikdel, K. and Firoozjai, M.N. (2014). Effect of different concentrations of IBA and time of taking cutting on rooting, growth and survival of *Ficus binnendijkii* Amstel Queen' cuttings. *Not. Sci. Biol.*, **6**(2), 163-166. doi: <https://doi.org/10.15835/nsb629281>
- Czaja, E. A. R., Moreira, R. R., Rozwalka, L. C., Figueiredo, J. A. G. and Mio, L. L. M. D. (2016). Gray mold in immature fig fruit: Pathogenicity and growth temperature. *Cienc. Rural.*, **46**, 1524-1527.
- Darwesh, H. Y., Bazaid, S. A. and Samra, B. N. A. (2014). In vitro propagation method of *Ficus carica* at Taif governorate using tissue culture technique. *Int. J. Adv. Res.*, **2**(6), 756-761.
- Fogaca, C. M. and Fett-Neto, A. G. (2005). Role of auxin and its modulators in the adventitious rooting of Eucalyptus species differing in recalcitrance. *Plant Growth Regul.*, **45**(1), 1-10.
- Ghosh, A., Dey, K., Mani, A., Bauri, F. K. and Mishra, D. K. (2017). Efficacy of different levels of IBA and NAA on rooting of Phalsa (*Grewia asiatica* L.) cuttings. *Int. J. Chem. Stud.*, **5**(6), 567-571.
- Hartman, H., Kester, D., Davies, F. T., Geneve, R. L. and Wilson, S. B. (2017). *Plant Propagation: Principles and Practices* (9th ed.). Pearson.
- Husen, A., Iqbal, M., Siddiqui, S. N., Sohrab, S. S. and Masresha, G. (2017). Effect of indole-3-butyric acid on clonal propagation of mulberry (*Morus alba* L.) stem cuttings: Rooting and associated biochemical changes. *Biol. Sci.*, **87**(2), 161-166.
- Kaewchangwat, N., Thanayupong, E., Jarussophon, S., Niamnont, N., Yata, T., Prateepchinda, S. and Suttisintong, K. (2020). Coumarin-Caged compounds of 1-naphthaleneacetic acid as light-responsive controlled-release plant root stimulators. *J. Agri. Food Chem.*, **68**(23), 6268-6279.
- Khalid, W. K. and Ahmed, A. A. (2022). Study of some natural substances in rooting of two fig varieties. *Int. J. Agri. Stat. Sci.*, **18**(1), 183-188.
- Koyuncu, F. and Senel, E. (2003). Rooting of black mulberry (*Morus nigra* L.) hardwood cuttings. *J. Fruit Ornament Plant. Res.*, **11**(1-2), 125-130.
- Kumar, V., Radha, A. and Kumar, C. S. (1998). In vitro plant regeneration of fig (*Ficus carica* L. cv. Gular) using apical buds from mature trees. *Plant Cell Rep.*, **17**(8), 717-720.
- Kuntagol, P., Patil, D., Biradar, I., Nagaraja, M., Najjappanavar, A. and Pattepure, S. (2018). Effect of IBA & NAA on root parameters of hardwood cuttings of fig varieties. *Green Farming Int. J.*, **6**(6), 1064-1067.
- Lansky, E. P. and Paavilainen, H. M. (2010). *Figs: The genus Ficus*. CRC Press.
- Mewar, D. and Naithani, D. C. (2016). Effect of different IBA concentrations and planting time on stem cuttings of wild fig (*Ficus palmata* Forsk.). *Plant Arc.*, **16**(2), 959-962.
- Mohana, M., Majd, A., Jafari, S., Kiabi, S. and Paivandi, M. (2014). The effect of various concentrations of IBA and NAA on the rooting of semi-hardwood cuttings of *Azalea alexander*. *Adv. Environ. Biol.*, **8**(16), 2223-2231.
- Park, S. M., Won, E. J., Park, Y. G. and Jeong, B. R. (2011). Effects of node position, number of leaflets left, and light intensity during cutting propagation on rooting and subsequent growth of domestic roses. *Horticulture, Environ. Biotechnol.*, **52**(4), 339-343.
- Patel, H. R., Patel, M. J. and Singh, T. (2017). Effect of different levels of IBA and NAA on rooting of hardwood and semi-hardwood cutting in fig. *Int. J. Agri. Sci. Res.*, **7**(4), 519-523.
- Rahdari, P., Khosroabadi, M., Delfani, K. and Hoseini, S. M. (2014). Effect of different concentrations of plant hormones (IBA and NAA) on rooting and growth factors in root and stem cuttings of

- Cordyline terminalis*. *J. Med. Biol. Sci.*, **3(3)**, 105-110.
- Reddy, K. V., Reddy, C. P. and Goud, P. V. (2008). Effect of auxins on the rooting of fig (*Ficus carica* L.) hardwood and semi-hardwood cuttings. *Ind. J. Agri. Res.*, **42(1)**, 75-78.
- Shamsuddin, M. S., Shahari, R., Amri, C. N. A. C., Tajudin, N. S., Mispan, M. R., & Salleh, M. S. (2021). Early development of fig (*Ficus carica* L.) root and shoot using different propagation medium and cutting types. *Trop. Life Sci. Res.*, **32(1)**, 83-93.
- Singh, K. K., & Tomar, Y. K. (2015). Effect of planting time and indole butyric acid levels on rooting of woody cuttings of phalsa (*Grewia asiatica* L.). *Hort Flora Res. Spec.*, **4(1)**, 39-43.
- Tsipouridis, C., Thomidis, T. and Isaakidis, A. (2003). Rooting of peach hardwood and semi-hardwood cuttings. *Aust. J. Exp. Agri.*, **43(11)**, 1363-1368.
- Vora, J. D., Vora, D., Pednekar, S. R., Patwardhan, A. U. and Shaikh, S. (2017). Biochemical, organoleptic assessment of fig (*Ficus carica*). *J Biotech. Biochem.*, **3(2)**, 95-104.
- Yusnita, Y., Jamaludin, J., Agustiansyah, A. and Hapsoro, D. (2017). A combination of IBA and NAA resulted in better rooting and shoot sprouting than single auxin on Malay apple [*Syzygium malaccense* (L.) Merr. & Perry] stem cuttings. *J. Agri. Sci.*, **40(1)**, 80-90.