



SUSTENANCE OF SOIL MICROBIAL BIOMASS, THE BASIS OF SOIL FERTILITY IN THE AGRO-ECOSYSTEMS: INFLUENCE OF PESTICIDE AND SOIL AMENDMENTS

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Abstract

Intensive agriculture has raised global concerns and has stimulated scientists worldwide to devise suitable agricultural management practices in order to achieve sustained soil fertility which support productivity in long term. Pesticides effect on soil microbial biomass that plays a key role in driving many ecosystem processes in the soil ecosystem is often argued and no clear cut information is available till date. Maintenance of soil microbial biomass is essential to sustain soil fertility. We critically evaluate the current evidences for this argument. Soil fertility maintenance involves considerable intervention in the research agenda. Most of studies done with pesticides are from laboratories or green house experiments; however the field conditions are inconspicuously lacking. There are a few studies indicating towards the soil fertility sustenance when pesticides are applied in combination with the soil amendments. We give recommendations to test the many other soil amendments that may be applied with different pesticides to see the possible ecological implications leading to designing of new eco-technologies to maintain and sustain the soil fertility.

Key Words: pesticides, soil amendments, soil microbial biomass, soil fertility, ecosystem processes.

Introduction

Pesticides application has now become the main method to control diseases, insects and weeds that infest crops (Jardim and Caldas 2012). The tropics with favorable environmental conditions for insect pests and weeds to proliferate lead to a greater reliance on pesticides than in temperate regions recently along agricultural expansion and changing agricultural practices (Sanchez-Beyo and Hyne 2011; Scopel *et al.*, 2013). The impact of pesticides varies from being hazardous to the ecosystem, health, persistence in the environment or even bioaccumulate (EPA 2017). Concerns have developed, however over the long term sustainability and negative environmental consequences of the intensification of agricultural system (Ladha *et al.*, 2003; Mandal *et al.*, 2003; Moreno *et al.*, 2007).

The extensive use of pesticides in the agricultural system since 1950's has played a pivotal role in the sustenance of crops (López *et al.*, 2002; Cycon *et al.*, 2006; Yang *et al.*, 2007) and lowering of agricultural costs in terms of input of labor and energy involved in agricultural production (Ayansina 2009). These are composed of active and inert ingredients in their formulations. The byproducts of pesticides being in inactive form which are present in the soil can still pose

threat to non-target organisms leading to the environmental degradation (Candiotti *et al.*, 2010; Vieira *et al.*, 2014), pollution of air, water and soil globally (Abhilash *et al.*, 2012; Volchko *et al.*, 2014) and their residues affects the humans in turn (Abhilash and Singh 2009). Only 1% of the pesticide is utilized by the plants in the protection against pests and rest of it leaches into the soil (Chang *et al.*, 2013). And the excess quantity of pesticides not reaching to the target organisms is absorbed by the plants (Ahemad and Khan 2011).

Most of the pesticides after application during soil management practices eventually reach the soil and thus cause changes in the growth and activity of soil microbial biomass as well (Floch *et al.*, 2011; Kumar *et al.*, 2012) that pertains to both the soil health and fertility levels (Fig. 1). The soil microbial biomass exhibits specific roles in the soil ecosystem. It constitutes the primary soil decomposers, a vital part of soil food web and the main drivers of key ecosystem processes such as organic matter decomposition, nutrient cycling and, thereby plant productivity (Pandey and Singh 2004) which comprise the basis of soil fertility (Singh *et al.*, 2011). Pesticides addition in general affects the soil microbial biomass not only quantitatively but also qualitatively resulting in changes

in the soil biochemical process, the indicators of soil fertility and plant yield (Dutta *et al.*, 2010; Xu *et al.*, 2013).

Different soil management practices such as tillage, crop rotation, application of various agrochemicals, chemical fertilizers and organic resources have a profound influence on the size and activity of the soil microbial biomass (Kushwaha and Singh 2005; Singh and Ghoshal 2007; Singh *et al.*, 2016). Microbial biomass responds much more rapidly to management changes than does the organic matter as a whole and hence its measurement has been mostly used as an early indicator of changes in soil chemical and physical properties resulting from management and environmental stress in agroecosystems (Doran and Zeiss 2000). Plenty reports are available related to the extensive and excessive use of pesticides in soil and possible side effects on the soil microbes (Getenga *et al.*, 2000; Xie *et al.*, 2004; Gundi *et al.*, 2005; Moreno *et al.*, 2007). In realistic agricultural conditions the pesticides are applied in combination with exogenous soil amendments like chemical fertilizer, animal manure, green manure and crop residues etc. which may interact with each other within the soil systems (Singh and Ghoshal 2010). The addition of exogenous soil amendments is required to maintain the soil fertility level in general. The effect of a pesticide may be changed when it interacts with other soil amendments coexisting in the soil, and such changes would have different side-effects on the biological function of the soil. There is, therefore, an increasing concern on the behaviors of combined application of pesticides and exogenous soil amendments and their potential effects on soil quality (Briceno and Palma 2007; Singh and Ghoshal 2016). Many reviewers have indicated the effects of pesticides even at recommended dosage application on soil microorganisms (Chowdhury *et al.* 2008; Lo 2010). In croplands, maintenance of soil fertility for sustained high crop production is a challenging task. This review compiles the rich data accumulated in literature since the beginning of twenty first century revealing the effect of pesticides both singly and in combination with exogenous soil amendments on soil microbial biomass. The purpose of this review is to assess briefly the effects of the use of pesticides and to make an initiative towards the study of its effect along soil amendments on soil fertility leading to replacement of agronomy from the centre of agriculture.

Effect of Pesticides on Soil Microbial Biomass

Soil microbial biomass is involved in the decomposition of organic matter and thus, the nutrient cycling in soils. Although soil microbial C constitutes 1-

3% of the total soil organic C and soil microbial N up to 5% of the total organic N, they are the most labile C and N pools in soil (Jenkinsen and Ladd 1981). The turnover time for C and N immobilized into the microbial biomass has been reported to be about ten times faster than that derived from plant materials (Smith and Paul 1990). Soil microbial biomass acts as the repository of major nutrients in soil via immobilization while release nutrients in soil through mineralization for the growth of plants and is therefore considered an indicator of soil fertility (Marumoto 1984; Hassink *et al.*, 1991). Thus both the size and the activity of microbial biomass determine the nutrient availability and productivity of soils (Singh and Ghoshal 2010; Singh *et al.*, 2011). Soil microbial biomass is considered to be the primary factor in the maintenance of not only the soil health and yield but also the sustainability (Zhao *et al.*, 2014a). Therefore, the analysis of the impacts on soil microbial biomass as a result of adopted various soil management practices will help us in selecting the suitable management strategy for the establishment of more stable and sustainable agro-ecosystems (Li *et al.*, 2012; Zhao *et al.*, 2014b). The effect of pesticides on soil microbial biomass has been reported from negative (Busse *et al.*, 2001; Sofu *et al.*, 2012), positive (Moreno *et al.*, 2007; Das *et al.*, 2013) to no effect (Lupwayi *et al.*, 2004; Lupwayi *et al.*, 2007) but there is no clear cut information (Table 1).

Gomez *et al.* (2009) while studying the influence of prolonged use of glyphosate on soil microbial biomass C in a field trial at Zavala, Argentina being in use since 1997 with soybean cropping found that the soil microbial biomass C was significantly lower with dosage of 1.92 and 3.84 L a.i. ha⁻¹ while it was lower but comparable to that of control with dosage of 0.48 and 0.96 L a.i. ha⁻¹ after four days of application. Contradictory results were found after 45 days of application when the soil microbial biomass C was found higher in dosage of 0.48 L a.i. ha⁻¹ while comparable in 3.84 L a.i. ha⁻¹ as compared to control. This was explained on the basis that glyphosate can not only stimulate but also inhibit soil microorganisms, depending on the soil type and concentration of the herbicide use (Carlisle and Trevors 1986). Several researches had reported for glyphosate to act as the source of major nutrients like C, N and P in soil which is readily utilized by soil microbes (Dick and Quinn 1995; Busse *et al.*, 2001). Glyphosate, an organophosphate is readily used by various soil microbes like gram-positive or gram-negative bacteria (Van Eerd *et al.*, 2003), fungal population (Araujo *et al.*, 2003; Ratcliff *et al.*, 2006) and the actinomycetes (Araujo *et al.*, 2003) as it acts as a rich source of energy and nutrients leading to an increase in bacterial biomass

(Zabaloy *et al.*, 2008) and also the fungal and actinomycetes (Araujo *et al.*, 2003). However, no change in soil microbial biomass of forest soils was reported by Busse *et al.*, 2001. Glyphosate when degraded by soil microbes into aminomethyl fosfo'nic acid (AMPA) soon turn into water, carbon dioxide, ammonia and phosphate (Dick and Quinn 1995). Though glyphosate is easily degraded yet its half life ranges from days to months due to its association with the organo-mineral complex (Jonge and Jonge 1999). The glyphosate mineralization rate is related to the accumulation and activity of soil microbes which indicates its persistence in soil.

Several reports are available indicating that chlorpyrifos inhibited soil microbial populations like bacteria, fungi, and actinomycetes initially after its application (Pandey and Singh 2004; Shan *et al.*, 2006; Chu *et al.*, 2008). Contradictory to this chlorpyrifos had also been reported to stimulate the growth of soil bacteria and fungi (Pozo *et al.*, 1995; Pandey and Singh 2004; Shan *et al.*, 2006). Such variations in the effect of a pesticide on soil microbes is attributed to concentration of pesticide applied, soil type, and microbial composition in tested soil. The deleterious effect of chlorpyrifos on soil microbes was enhanced when applied in combination with another pesticide chlorothalonil which depended on its concentration (Chu *et al.* 2008). Both herbicides together inhibit acetolactate synthase thus having low and indirect effects on soil microbial activities (Perucci *et al.*, 1999).

Various studies are available depicting the negative effect of the herbicide bromoxynil, a sulfonyl urea on the soil microbial biomass (El-Ghamry *et al.*, 2000, Pampulha and Oliveira 2006, Abbas *et al.*, 2014). Allievi and Gigliotti (2001) explained reduction in soil microbial biomass on the basis of lowering in amino acid assimilation ability of bacteria which resulted in the death of bacteria and thereby the reduction in soil microbial biomass. Ratnayak and Audus (1987) found the decrease in nitrifying bacteria due to bromoxynil herbicide. Abbas *et al.*, 2014 explained on the basis of restricted hydrolysis at high pH leading to a higher persistence of herbicide resulting in the death of microbes and thus a reduction in soil microbial biomass.

Atrazine application is reported to exert significant changes in content of organic matter (Ayansina and Oso 2006). High organic matter content in soil is generally considered the prime factor among several factors like pesticides concentration, soil type etc. to be responsible for the persistence of any pesticide. Increase in duration of exposure of soil microbial biomass to pesticide resulted in decline in the content of soil microbial

biomass. On contrary, Das *et al.* (2012) also found the increase in soil microbial biomass up to the 45 days of application of pendimethalin with fenoxaprop/ paraquat. Das and Debnath (2006) reported the stimulatory effect of herbicides on not only the quantity but also the function of non-symbiotic N₂-fixing bacteria and phosphate-solubilizing microorganisms respectively. The higher accumulation of soil microbial biomass in the herbicide treated soils in these studies revealed that the herbicides along their fractions and the autolyzed dead cells were used by the soil microbes to get the C, energy and other nutrients required for the growth and metabolism of soil microbes (Das *et al.*, 2012; Perucci and Scarponi 1994). Similarly, Nongthombam *et al.* (2008) while studying the comparative residual effects of widely used aryl phenoxy propionic acid, dinitroaniline and bipyridilium herbicides on microbial biomass and availability of plant nutrients under a particular set of soil conditions found that the herbicides are degraded by the soil microbes when the catabolised/ co-metabolized fractions of herbicides remain in soil as oxidizable C. The negative effect of pesticides on soil microbial biomass decreased with time as the pesticide was degraded or the microbes became adapted to these agrochemicals. One another reason behind the increase in soil microbial biomass after a decrease for a short span of time was the multiplication of microbes due to increase in nutrients in soil or killing of microbes by the pesticides (Latha and Gopal 2010; Vandana *et al.*, 2012). The decrease in soil microbial biomass depends upon the variability in the type, dosage, rate of application of pesticides (Ayansina and Oso 2006; Sebiomo *et al.*, 2011), mode of application, group of microorganisms and the environmental conditions (Subhani *et al.*, 2000; Zain *et al.*, 2013). The deleterious effect of pesticides on soil microbial biomass may also be attributed to the lower input of organic residues (Wainwright 1978).

The decrease in soil microbial biomass caused by the application of butachlor at recommended rate (Singh and Ghoshal 2010) was attributed to the lowering in fungal biomass as compared to the other soil microbes (Min *et al.*, 2007; Xia *et al.*, 2012). However, the higher soil microbial biomass due to butachlor application was on account of its degradation, while the higher concentration of butachlor resulted in the lowering in the soil microbial biomass (Xia *et al.*, 2012).

No changes in soil microbial biomass have been found in studies done by Lupwayi *et al.* (2009) due to the effect of application of herbicides at the recommended rate although the functional structures of bacterial communities were altered. Lupwayi *et al.* (2010) in a field experiment with canola-barley crop rotation while studying the effect of 100% of the

recommended herbicide rates viz., tralkoxydim at 200 g ai ha⁻¹ and bromoxynil + MCPA at 560 g ai ha⁻¹ on the soil microbial biomass found that it was significantly decreased when was applied at full rates while no effect was seen at 50 % rate of application. They suggested that the deleterious effect of herbicide on soil microbial biomass was reduced on reducing the application rate. This study also implied that herbicide when applied at recommended rate in single studies did not show any effect however the application of herbicides for a long duration could result in decreasing the soil microbial biomass.

Assessment of the effect of pesticides on soil microbial biomass is difficult to understand in soil due to different research findings as reported in the literature. A number of factors could be responsible for those controversial results such as soil properties, chemical nature and concentration of pesticides, biological function observed. Even if pesticides applied at recommended rates may cause slight and transient changes to populations or activities of soil microorganisms (Johnsen *et al.* 2001), it is obvious that long-term recurrent applications of pesticides are known to interfere with the biochemical balance, which can reduce soil fertility and productivity by affecting local metabolism. To preserve the environment, many of those molecules have been and will be withdrawn from the market such as clothianidine, imidaclopride, thiamethoxam and endosulfan or methods be devised to overcome the side effects of pesticides.

The synthesis of the review is reported in Fig. 2. It highlights the effect of pesticides from 44 studies done in twentieth century as depicted in this review varies from positive to negative to no effect on soil microbial biomass which is generally represented as the early indicator of soil fertility. The variability in responses of soil microbial biomass depends on the concentration of pesticides applied, rate of application, soil type, environmental conditions etc. The general trends highlighted and reported in Fig. 2 have been compared with numerous results from other articles dealing with the impact of pesticides on soil microbial biomass. Moreover the response of soil microbial biomass varies with the pesticides dosage, number of times of application, agricultural condition etc. This review attempts to identify common determinants explaining variation in patterns of soil microbial biomass in relationship with application of different types of pesticides aiming at understanding the impact of pesticides on soil microbes. The deleterious effect on soil microbial biomass due to application of pesticides may further be changed when applied in combination with soil amendments of varying resource quality. The studies revealing deleterious effects of pesticides on soil

fertility indices demands an urge to establish such agricultural practice which not only maintain the soil fertility level but also sustain it for long term. This perhaps led to the application of pesticides along soil amendments.

Effect of Pesticides in Combination with Soil Amendments on Soil Microbial Biomass:

Most of the researches have been done with a single application of pesticides (Lupwayi *et al.*, 2007; Vischetti *et al.*, 2007). The influence of pesticides on soil microbial biomass is attributed to its toxic behavior and their fate in soil to which many processes like adsorption, leaching, run-off, degradation, volatilization, plant uptake, etc. contribute (Jacobsen and Hjelmsø 2014). The pesticide content remaining after such processes is considered bioavailable to affect the soil microbial biomass which is further modified by the application of soil amendments (Herrero-Hernández *et al.*, 2011).

Soil amendment is the process of adding manure, fertilizers, vermicompost, sewage sludge, wheat straw to the dry land covering upto 80% of global area which not only increases the organic matter, moisture content of the soil but also enhances the nutrient level necessary for the growth of the plant and crop yield. (Bastida *et al.*, 2015; Singh *et al.*, 2016; Singh and Ghoshal 2010; Pose-Juan *et al.*, 2017). The nutrient limited condition favors growth and accumulation of k-strategists compared to r-strategists which accumulate in conditions of readily mineralizable C and N in soil. When pesticides are applied in combination with soil amendments, interaction may occur resulting in the changes in the efficacy of pesticide. The interaction between both amendments and pesticides may further result in the changes in levels of soil microbial biomass.

Pose-Juan *et al.* (2017) reported increased soil microbial biomass carbon while studying the combined effect of triasulfuron herbicide at the rate of 10 and 50 mgkg⁻¹ along green compost and at the rate of 2 mg kg⁻¹ along sewage sludge. Similarly, Singh *et al.* (2016) while studying the effect of herbicide butachlor singly and in combination with soil amendments in a rice, wheat, summer fallow tropical dryland agroecosystem found significantly higher soil microbial biomass C, N and P throughout the annual cycle in combined treatments compared to control while the increase in the herbicide only treatment was comparable to that of control. The increase found was justified on the basis of the resource quality of soil amendments which masked the effect of herbicide when applied in combination. They found the higher levels of soil microbial biomass C, N and P in case of soil amended with butachlor and *Sesbania* in combination during early phase of annual

cycle and explained this on the basis of high nutrient availability owing to its faster decomposition on account of lower C:N ratio. This same treatment showed lower soil microbial biomass during the later phase of annual cycle which was due to the lower availability of nutrients for either plant uptake or to be utilized by soil microbes.

Prolonged higher levels of soil microbial biomass were found in case of farmyard manure application along butachlor herbicide in the studies conducted by Singh and Ghoshal 2010 and Singh *et al.* 2016 and was explained by the fact that there are different fractions in farmyard manure which decompose at different rates and thus maintaining the sustained availability of nutrients (Sluijsmans and Kolenbrander 1977). Higher levels of soil microbial biomass C, N and P was also found by Das *et al.* (2015) while studying the effect of thiobencarb, pendimethalin and pretilachlor at the rate of 7.5, 10 and 2.5 kg a.i.ha⁻¹ when applied in soil amended with farmyard manure. On contrary lower soil microbial biomass carbon was reported by Perucci *et al.* (2000) on application of herbicides rimsulfuron and imazethapyr at both the recommended rate of application and ten times the recommended rate of application when applied in combination with vermicompost.

Lower soil microbial biomass during early phase of annual cycle while higher during later phase of annual cycle was observed in case of herbicide applied along wheat straw and was explained on the basis of low resource quality of wheat straw i.e., the lower C:N ratio. Soil amendments with low resource quality is reported to immobilize nutrients during early phase which get remineralized during a later phase of the annual cycle leading to higher soil microbial biomass during the later phase of the annual cycle in a two year field study (Singh *et al.*, 2016; Singh and Ghoshal 2010). These studies gave clear evidence that the variation in soil microbial biomass was mainly due to the application of organic amendments rather than the herbicide in all the combined applications.

Lupwayi *et al.* (2010) conducted a four year field experiment to compare the effect of chemical fertilizer (NPKS) at recommended rate along 100% of the recommended dose of herbicide viz., glufosinate-ammonium at 500 g a.i. ha⁻¹ and clethodim at 30 g a.i. ha⁻¹, high yielding variety of crop, optimum seeding

named as full package, full – 50% of the recommended rates of fertilizer and full – 50% of pesticides in canola and tralkoxydim and bromoxynil + MCPA in barley during canola - barley crop rotation on the response of soil microbial biomass. They found that the pesticides application had a negative effect on soil microbial biomass C which was limited in the treatment full – 50% pesticide.

Higher soil microbial biomass on account of application of pesticides in combination with soil amendments (Singh *et al.*, 2016) can stabilize the soil ecosystem (Chauhan *et al.*, 2006) due to their ability to contribute to several ecosystem processes and to enhance the plant productivity in turn.

Conclusion and Future Perspectives

The addition of pesticides in modern agriculture lead to the serious consequences on the environmental resources which further affects the health of soil in terms of soil microbial biomass. An overview of the data presented in this review article highlights the variations in the responses of soil microbial biomass due to application of different pesticides singly and in combination with soil amendments. Hence, it is the need of the hour to devise the suitable environmental friendly agricultural strategies aiming at reducing the negative effect of pesticides on the soil microbial biomass. A summary of findings in the present review indicated that diverse soil amendments when applied in combination might be involved in ameliorating the negative impact of pesticides. However, the diversity of contexts and approaches that constitute the basis of this analysis can strengthen our conclusions. Thus the application of pesticides in combination with soil amendments of varying resource quality could be useful for researchers and for policy decision makers in order to take the place of agronomy on which agriculture is generally focused.

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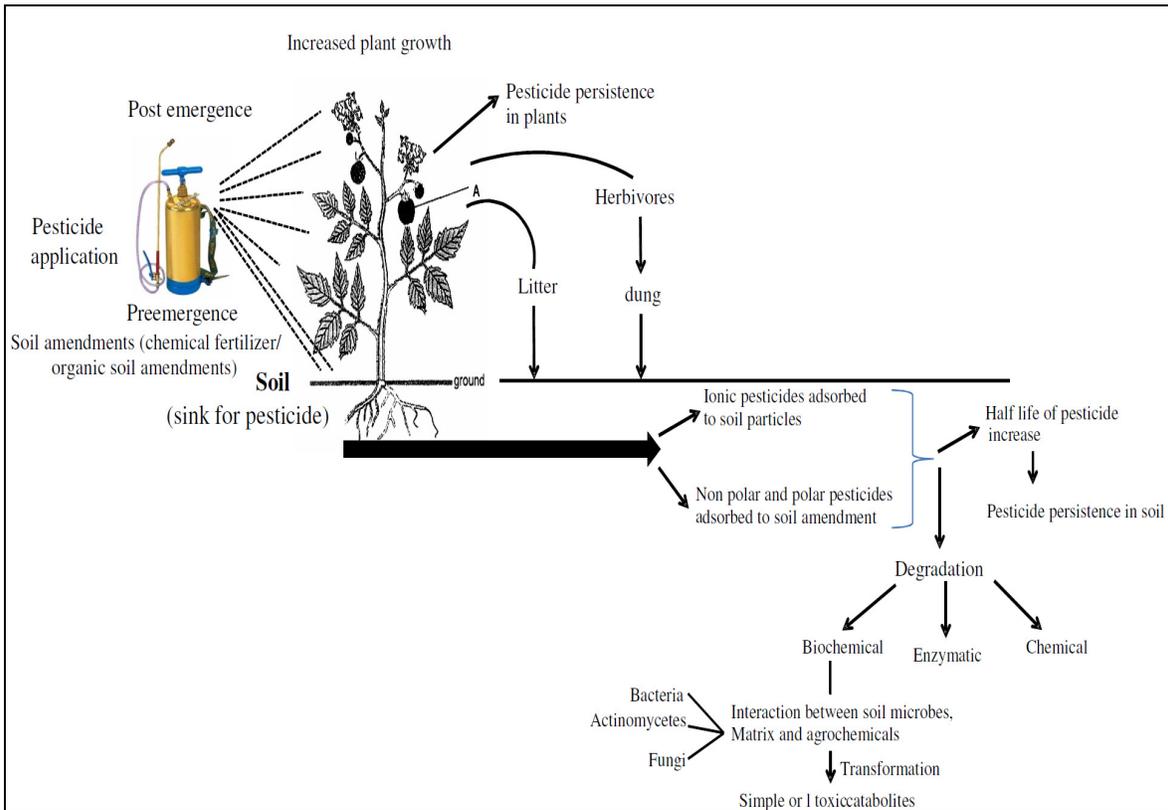


Fig. 1 : Schematic representation of pesticides accumulation in agroecosystems.

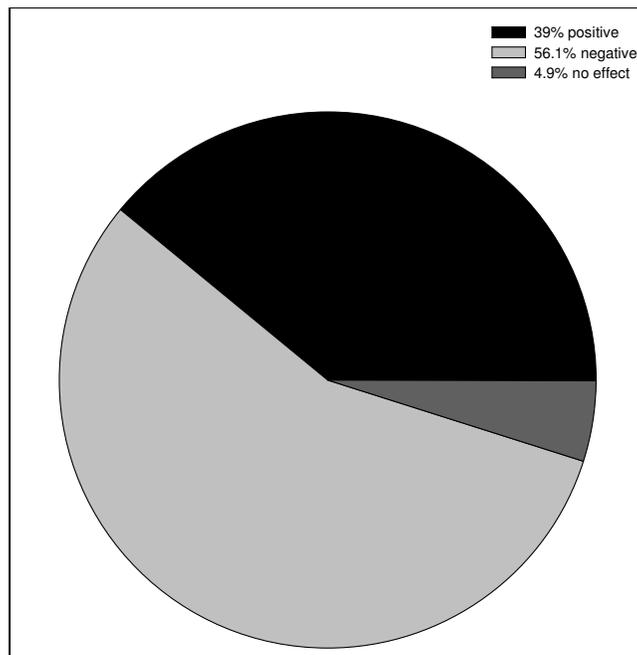


Fig. 2 : Response of soil microbial biomass to various pesticides from the total forty one studies reported in this review.

Table 1 Effect of application of pesticides on soil microbial biomass

S No.	Experimental conditions	Pesticide	Class	Dosage	Mode of application	Effect	References
1.	Ponderosa Plantations	Glyphosate	Miscellaneous	900ga.i.ha ⁻¹	post emergence	no effect	Busse et al. 2000
2.	Laboratory cond.	Rimsulfuron	Sulfonylurea	fr,10fr, 100fr		transient decrease in SMBC	Vischetti et al. 2000
3.	Laboratory cond.	Imazamox	Imidazolinone	50% dose		20% decrease in SMBC	Vischetti et al. 2002
4.	Laboratory cond.	Chlorsulfuron	Triazenc	fr,10fr, 100fr		decrease in SMBC	Ghamry et al. 2000
5.	Laboratory cond.	Rimsulfuron Imazethapyr	Sulfonylurea Imidazolinones	fr. 10fr		decrease in SMBC	Perucci et al. 2000
6.	Laboratory cond.	Glyphosate Glufosinate-ammonium Sethoxydim Bentazon azoles Imazamet-habenz Dicamba Clopyralid acids 2, 4-D amine Metribuzin Imazamox/ Imazethapyr Triasulfuron Metsulfuron-methyl	Miscellaneous Miscellaneous Cyclohexane-diones Benzothiad- Imidazolinones Benzoic acids Carboxylic Phenoxys Triazinones Imidazolinones Sulfonylureas Sulfonylureas	900ga.i.ha ⁻¹ 500ga.i.ha ⁻¹ 200ga.i.ha ⁻¹ 1100ga.i.ha ⁻¹ 400ga.i.ha ⁻¹ 110ga.i.ha ⁻¹ 200ga.i.ha ⁻¹ 560ga.i.ha ⁻¹ 210ga.i.ha ⁻¹ 30ga.i.ha ⁻¹ 10ga.i.ha ⁻¹ 4.5ga.i.ha ⁻¹	all post emergence	no effect	Lupwayi et al. 2004
7.	Field condition	Glyphosate	Miscellaneous	900ga.i.ha ⁻¹	post emergence	no effect	Lupwayi et al. 2007
8.	Italian bilobed	Chlorpyrifos	Organophosphate	10mg/kg 50mg/kg		decrease by 25% and 50%	Vischetti et al. 2007
9.	Laboratory cond.	2, 4-D	Phenoxys	10mg/kg		decrease in SMBC	Macur et al. 2007
10.	Laboratory cond.	atrazine	triazine	0.2 to 1000 mg/kg		increase in SMB	Moreno et al. 2007
11.	Laboratory cond.	Triasulfuron	Sulfonylureas	10 fr		decrease SMB	Sofo et al. 2012
12.	Field experiment	butachlor, paraquat glyphosate	chloroacetanilide pyrozosulfuran, bipyridilium organophosphate	1 kg/ha 25g/ha 200 g/l 360 g/l	post emergence	decrease in MBC " " "	Baboo et al. 2013
13.	Laboratory cond.	Fenoxaprop pendimethalin paraquat	arylphenoxy-propionic acid dinitroaniline bipyridilium	50ga.i.ha ⁻¹ 1kga.i.ha ⁻¹ 1kga.i.ha ⁻¹		39.8% inc. in MBC 37.1% inc. in MBN, 28.2% inc. in MBC 15.2% inc. in MBP	Das et al. 2013
14.	Field conditions	Butachlor	Chloroacetanilide	2 kga.i. ha ⁻¹	pre emergence	no effect on SMBC, SMBN and SMBP	Singh et al. 2016
15.	Laboratory cond.	Triasulfuron	Sulfonylureas	2, 10 and 50 mgkg ⁻¹		decreased SMBC	Pose-Juan et al. 2017
16.	Field condition	Imidacloprid	Neonicotinoid	fr. 2fr, 5fr, 10fr	post emergence	Transient decrease in SMBC	Mahapatra et al., 2017

SMBC: Soil microbial biomass carbon, SMBN: Soil microbial biomass N, SMBP: Soil microbial biomass P, cond.: condition, fr: recommended rate

References

- Abbas, Z.; Akmal, M.; Khan, K.S. and Hassan, F. (2014). Effect of buctril super (bromoxynil) herbicide on soil microbial biomass and bacterial population. *Braz. Arch. Biol. Tech.* 57 : 9–14.
- Abhilash, P.C. and Singh, N. (2009). Pesticide use and application: an Indian scenario. *J. Haz. Mat.* 165 : 1–12.
- Abhilash, P.C.; Powell, J.; Singh, H.B. and Singh, B. (2012). Plant-microbe interactions: novel applications for exploitation in multipurpose remediation technologies. *Trends Biotech.* 20: 416–420.
- Ahemad, M. and Khan, M.S. (2010). Comparative toxicity of selected insecticides to pea plants and growth promotion in response to insecticide-tolerant and plant growth promoting *Rhizobium leguminosarum*. *Crop Prot.* 29 : 325–329.
- Ahemad, M. and Khan, M.S. (2011). Pesticide interactions with soil microflora: importance in bioremediation. In: Ahmad I., Ahmad F., Pichtel J. (Eds) *Microbes and Microbial Technology: Agricultural and Environmental Applications*. Springer, New York, 393–413.
- Allievi, L. and Gigliotti, C. (2001). Response of the bacteria and fungi of two soils to the sulfonylurea herbicide cinosulfuron. *J. Env. Sci. Health B.*, 36 : 161–175.
- Araújo, A.S.F.; Monteiro, R.T.R. and Abarkeli, R.B. (2003). Effect of glyphosate on the microbial activity of two Brazilian soils. *Chemosphere* 52 : 799–804.
- Ayansina, A.D.V. (2009). Pesticide use in agriculture and microorganisms. In: M.S. Khan, A. Zaidi, and J. Musarrat (Eds) *Microbes in Sustainable Agriculture*. Nova Science Publishers Inc, New York, pp. 261–284.
- Ayansina, A.D. and Oso, B.A. (2006). Effect of two commonly used herbicides on soil microflora at two different concentrations. *Af. J. Biotech.*, 5 : 129–132.
- Biederbeck, V.O.; Campbell, C.A. and Smith, A.E. (1987). Effects of long-term 2, 4-D application on soil biochemical processes. *J. Environ. Qual.* 16 : 257–262.
- Bastida, F.; Selevsek, N.; Torres, I.F.; Hernández, T. and García, C. (2015). Soil restoration with organic amendments: linking cellular functionality and ecosystem processes. *Sc. Rep.*, 5 : 15550.
- Busse, M.D.; Ratcliffe, A.W.; Shestak, C.J. and Powers, R.F. (2001). Glyphosate toxicity and the effects of long-term vegetation control on soil microbial communities. *Soil Biol. Biochem.*, 33 : 1777–1789.
- Briceno, G. and Palma, G. (2007). Influence of Organic Amendment on the Biodegradation and Movement of Pesticides. *Cr. Rev. Environ. Sc. Tech.*, 37 : 233–271.
- Candiotti, J.V.; Natale, G.S.; Soloneskia, S.; Ronco, A.E. and Larramendy, M.L. (2010). Sublethal and lethal effects on *Rhinella arenarum* (Anura, Bufonidae) tadpoles exerted by the pirimicarb-containing technical formulation insecticide Aficida®. *Chemosphere* 78 : 249–255.
- Carlisle, S.M. and Trevors, J.T. (1986). Effect of the herbicide glyphosate on respiration and hydrogen consumption in soil. *Water Air Soil Poll.*, 27 : 391–401.
- Chang M., Wang, M.; Kuo, D.T.F. and Shih, Y. (2013). Sorption of selected aromatic compounds by vegetables. *Ecol. Eng.* 61 : 74–81.
- Chauhan, A.K.; Das, A.; Kharkwal, H.; Kharkwal, A.C. and Varma, A. (2006). Impact of microorganisms on environment and health. In: A.K. Chauhan and A. Varma (Eds) *Microbes: Health and environment*. IK International Publication House, New Delhi, India, pp. 1–12.
- Chowdhury, A.; Pradhan, S.; Saha, M. and Sanyal, N. (2008). Impact of pesticides on soil microbiological parameters and possible bioremediation strategies. I. *J. Microbiol.* 48 : 114–127.
- Chu, X.; Fang, H.; Pan, X.; Wang, X.; Shan, M.; Feng, B. and Yu, Y. (2008). Degradation of chlorpyrifos alone and in combination with chlorothalonil and their effects on soil microbial populations. *J. Environ. Sc.*, 20 : 464–469.
- Cycon, M.; Piotrowska-Seget, Z.; Kaczynska, A. and Kozdro, J. (2006). Microbiological characteristics of a sandy loam soil exposed to tebuconazole and l-cyhalothrin under laboratory conditions. *Ecotoxicology*, 15 : 639–646.
- Das, A.C. and Debnath, A. (2006). Effect of systemic herbicides on N₂-fixing and phosphate solubilizing microorganisms in relation to availability of nitrogen and phosphorus in paddy soils of West Bengal. *Chemosphere*, 65 : 1082–1086.
- Das, A.C.; Nayek, H. and Nongthombam, S.D. (2012). Effect of pendimethalin and quizalofop on N₂-fixing bacteria in relation to availability of nitrogen in a Typic Haplustep soil of West Bengal, India. *Environ. Mon. Assess.*, 184 : 1985–1989.
- Das, A.C.; Barman, S. and Das, R. (2015). Effect of pre-emergence herbicides on microbial biomass and biochemical processes in a typic fluvaquent soil

- amended with farmyard manure. *Bull. Environ. Cont. Tox.*, 95 : 395–400.
- Dick, R.E. and Quinn, J.P. (1995). Glyphosate-degrading isolates from environmental samples: occurrence and pathways of degradation. *App. Microbiol. Biotech.*, 43 : 545–550.
- Doran, J.W. and Zeiss, M.R. (2000). Soil health and sustainability: managing the biotic component of soil quality. *App. Soil Ecol.*, 15 : 3–11.
- Dutta, M.; Sardar, D.; Pal, R. and Kole, R.K. (2010). Effect of chlorpyrifos on microbial biomass and activities in tropical clay loam soil. *Environ. Monitor. Assess.*, 160 : 385–391.
- Fenner, K.; Canonica, S.; Wackett, L.P. and Elsner, M. (2013). Evaluating pesticide degradation in the environment: blind spots and emerging opportunities. *Science*, 341 : 752–758.
- Floch, C.; Chevremont, A.; Joanico, K.; Capowiez, Y. and Criquet, S. (2011). Indicators of pesticide contamination: Soil enzyme compared to functional diversity of bacterial communities via Biolog® Ecoplates. *Eur. J. Soil Biol.*, 47 : 256–263.
- Fontaine, S.; Mariotti, A. and Abbadie, L. (2003). The priming effect of organic matter: a question of microbial competition? *Soil Biol Biochem.*, 35 : 837–843.
- Getenga, Z.M., Jondiko, J.I.O.; Wandiga, S.O. and Beck, E. (2000). Dissipation behavior of malathion and dimethoate residues from the soil and their uptake by garden pea (*Pisum sativum*). *Bull. Environ. Cont. Toxicol.*, 64 : 359–367.
- El-Ghamry, A.M.; Chang-yong, H. and Jian-ming, X. (2000). Influence of chlorsulfuran herbicide on size of microbial biomass in soil. *J. Environ. Sc.*, 12 : 138–143.
- EPA. United States Environmental Protection Agency. Available at <http://www.epa.gov/pesticides>. Accessed 01 Aug 2017
- Gomez, E.; Laura, F.; Lorena, L. and Estela, F. (2009). Impact of glyphosate application on microbial biomass and metabolic activity in a Vertic Argiudoll from Argentina. *Eur. J. Soil. Biol.*, 45 : 163–167.
- Grenni, P.; Barra, C.A.; Rodríguez-Cruz, M.S. and Sánchez-Martín, M.J. (2009). Changes in the microbial activity in a soil amended with oak and pine residues and treated with linuron herbicide. *App. Soil Ecol.*, 41 : 2–7.
- Gundi, V.A.; Narasimha, G. and Reddy, B.R. (2005). Interaction effects of insecticides on microbial populations and dehydrogenase activity in a black clay soil. *J. Env. Sci. Health B.*, 40 : 269–283.
- Hassink, J.; Lebbink, G. and VanVeen, J.A. (1991). Microbial biomass and activity of a reclaimed-polder soil under a conventional or a reduced-input farming system. *Soil. Biol. Biochem.*, 23 : 507–513.
- Herrero-Hernández, E.; Andrades, M.S.; Marín-Benito, J.M.; Sánchez-Martín, M.J. and Rodríguez-Cruz, M.S. (2011). Field-scale dissipation of tebuconazole in a vineyard soil amended with spent mushroom substrate and its potential environmental impact. *Ecotoxic. Environ. Safety.*, 74 : 1480–1488.
- Jardim, A.N.O. and Caldas, E.D. (2012). Brazilian monitoring programs for pesticide residues in food e results from 2001 to 2010. *Food Control.*, 25 : 607
- Jacobsen, C.S. and Hjelmsø, M.H. (2014). Agricultural soils, pesticides and microbial diversity. *Curr. Opinion Biotech.*, 27 : 15–20.
- Jenkinson, D.S. and Ladd, J.N. (1981). Microbial Biomass in Soil: Measurement and Turnover. In: E.A. Paul and J.N. Ladd (Eds) *Soil Biochemistry*. Marcel Dekker, New York, 415–471.
- Johnsen, K.; Jacobsen, C.S.; Torsvik, V. and Sorensen, J. (2001). Pesticide effects on bacterial diversity in agricultural soils: a review. *Biol. Fert. Soils.*, 33 : 443–453.
- Jonge, H. and Jonge, L.W. (1999). Influence of pH and solution composition on the sorption of glyphosate and prochloraz to a sandy loam soil. *Chemosphere*, 39: 753–763.
- Pose-Juan, E.; Igual, J.M.; Curto, N.; Sánchez-Martín, M.J. and Rodríguez-Cruz, M.S. (2015a). Mesotrione dissipation and response of soil microbial communities in a soil amended with organic residues. *Spanish J. Soil Sc.*, 1: 12–25.
- Pose-Juan, E.; Sánchez-Martín, M.J., Herrero-Hernández, E. and Rodríguez-Cruz, M.S. (2015b). Application of mesotrione at different doses in an amended soil: dissipation and effect on the soil microbial biomass and activity. *Sc. Tot. Environ.*, 536 : 31–38.
- Pose-Juan, E.; Igual, J.M.; Sánchez-Martín, M.J. and Rodríguez-Cruz, M.S. (2017). Influence of Herbicide Triasulfuron on Soil Microbial Community in an Unamended Soil and a Soil Amended with Organic Residues. *Frontiers Microbiol.*, 8 : 378.
- Kumar, A.; Nayak, A.K.; Shukla, A.K.; Panda, B.B.; Raja, R.; Shahid, M.; Tripathi, R.; Mohanty, S. and Rath, P.C. (2012). Microbial biomass and carbon mineralization in agricultural soils as affected by pesticide addition. *Bull. Environ. Contam. Toxic.*, 88 : 538–542.
- Kushwaha, C.P. and Singh, K.P. (2005). Crop productivity and soil fertility in a tropical dryland

- agro-ecosystem: Impact of residue and tillage management. *Exp. Agric.*, 41 : 39–50.
- Ladha, J.K.; Pathak, H.; Tirol-Padre, A.; Dawe, D. and Gupta, R.K. (2003). Productivity trends in intensive rice–wheat cropping systems in Asia.” In: Ladha J. K. (Ed.) *Improving the Productivity and Sustainability of Rice–Wheat Systems: Issues and Impacts*. ASA, CSSA, and SSSA, Madison, WI, 45–76.
- Latha, P.C. and Gopal, H. (2010). Effect of herbicides on soil microorganisms. *Indian J. Weed Sc.*, 42 : 217–222.
- Lo Chi-Chu (2010). Effect of pesticides on soil microbial community. *J. Environ. Sc. Health B*, 45 : 348–359.
- Lo’pez, L.; Pozo, C.; Go’mez, M.A.; Calvo, C. and Gonza’les, J.L. (2002). Studies on the effects of the insecticide aldrin on aquatic microbial populations. *Int. Biodeterior. & Biodegrad*, 50 : 83–87.
- Li, R.; Khafipour, E.; Krause, D.O.; Entz, M.H.; deKievit, T.R. and Fernando, W.G.D. (2012). Pyrosequencing reveals the influence of organic and conventional farming systems on bacterial communities. *PLoS One*, 7 : e51897.
- Lupwayi, N.Z.; Harker, K.N.; Clayton, G.W.; Turkington, T.K.; Rice, W.A. and O’Donovan, J.T. (2004). Soil microbial biomass and diversity after herbicide application. *Can. J. Plant Sc.*, 84: 677–685.
- Lupwayi, N.Z.; Hanson, K.G.; Harker, K.N.; Clayton, G.W.; Blackshaw, R.E.; O’Donovan, J.T.; Johnson, E.N.; Gan, Y.; Irvine, B. and Monreal, M.A. (2007). Soil microbial biomass, functional diversity and enzyme activity in glyphosate-resistant wheat-canola rotations under low-disturbance direct seeding and conventional tillage. *Soil Biol. Biochem.*, 39 : 1418–1427.
- Lupwayi, N.Z.; Harker, K.N.; Clayton, G.W.; O’Donovan, J.T. and Blackshaw, R.E. (2009). Soil microbial response to herbicides applied to glyphosate-resistant canola. *Agric. Ecosys. & Environ.*, 129 : 171–176.
- Lupwayi, N.Z.; Stewart, A.B.; Harker, K.N.; O’Donovan, J.T.; Clayton, G.W. and Turkington, T.K. (2010). Contrasting soil microbial responses to fertilizers and herbicides in a canola - barley rotation. *Soil Biol. Biochem.*, 42 : 1997–2004.
- Macur, R.E.; Wheeler, J.T.; Burr, M.D. and Inskip, W.P. (2007). Impacts of 2, 4-D application on soil microbial community structure and on populations associated with 2, 4-D degradation. *Microbiol. Res.*, 162 : 37–45.
- Mahapatra, B.; Adak, T.; Patil, N.K.B.; Pandi, G.; Gowda, G.B.; Jambhulkar, N.N.; Yadav, M.K.; Panneerselvam, P.; Kumar, U.; Munda, S. and Jena, M. (2017). Imidacloprid application changes microbial dynamics and enzymes in rice soil Ecotoxic. *Environ. Safety.*, 144 : 123–130.
- Mandal, U.K.; Singh, G.; Victor. U.S. and Sharma, K.L. (2003). Green manuring: its effect on soil properties and crop growth under rice–wheat cropping system. *Eur. J. Agron.*, 19 : 225–237.
- Mandal, A., Patra, A.K.; Singh, D.; Swarup, A. and Ebhin Masto, R. (2007). Effect of long-term application of manure and fertilizer on biological and biochemical activities in soil during crop development stages. *Biores. Tech.*, 98 : 3585–3592.
- Marumoto, T. (1984). Mineralization of C and N from microbial biomass in paddy soil. *Plant Soil*, 76 : 165–173.
- Min, H.; Ye, Y.F.; Chen, Z.; Wu, W. and Yufeng, D. (2007). Effects of butachlor on microbial populations and enzyme activities in paddy soil. *J. Environ. Sc. Health B.*, 36 : 581–595.
- Moreno, J.L.; Aliaga, A.; Navarro, S.; Hernandez, T. and Garcia, C. (2007). Effects of atrazine on microbial activity in semiarid soil. *App. Soil Ecol.*, 35 : 120–127.
- Nongthombam, S.D.; Nayek, H. and Das, A.C. (2008). Effect of anilofos and pendimethalin herbicides on N₂-fixing and phosphate solubilizing microorganisms in relation to availability of nitrogen and phosphorus in a Typic Haplustep soil. *J. Crop Weed.*, 4 : 1–6.
- Omar, S.A. and Abdel-Sater, M.A. (2001). Microbial populations and enzyme activities in soil treated with pesticides. *Water Air Soil Poll.*, 127 : 49–63.
- Pandey, S. and Singh, D.K. (2004). Total bacterial and fungal population after chlorpyrifos and quinalphos treatments in groundnut (*Arachis hypogaea* L.) soil. *Chemosphere*, 55 : 197–205.
- Pampulha, M.E. and Oliveira, A. (2006). Impact of an herbicide combination of bromoxynil and prosulfuron on soil microorganisms. *Curr. Microbiol.* 53 : 238–243.
- Perucci, P.; Dumontet, S.; Bufo, S.A.; Mazaatura, A. and Casucci, C. (2000). Effects of organic amendment and herbicide treatment on soil microbial biomass. *Biol. Fertil. Soils*, 32 : 17–23.
- Perucci, P. and Scarponi, L. (1994). Effect of the herbicide imazethapyr on soil microbial biomass and various soil enzyme activities. *Biol. Fertil. Soils*, 17 : 237–240.
- Perucci, P.; Vischetti, C. and Battistoni, F. (1999). Rimsulfuron in a silt clay loam soil: Effects upon microbiological and biochemical properties under varying microcosm conditions. *Soil Biol. Biochem.*, 31 : 195–204.

- Pozo, C.; Martínez-Toledo, M.V.; Salmeron, S.; Rodelas, B. and Gonzalez-Lopez, J. (1995). Effect of chlorpyrifos on soil microbial activity. *Environ. Toxic. Chem.*, 14 : 187–192.
- Ratcliff, A.W.; Busse, M.D. and Shestak, C.J. (2006). Changes in microbial community structure following herbicide (glyphosate) additions to forest soils. *App. Soil Ecol.*, 34 : 114–124.
- Ratnayake, M. and Audus, L.J. (1978). Studies on the effects of herbicides on soil nitrification. II. *Pesticide Biochem. Phy.*, 8 : 170–185.
- Rodríguez-Liévana, J.A.; ElGouzi, S.; Mingorance, M.D.; Castillo, A.; Peña, A. (2014). Irrigation of a Mediterranean soil under field conditions with urban waste water: effect on pesticide behavior. *Agric. Ecosys. Environ.*, 185 : 176–185.
- Sanchez-Bayo, F. and Hyne, R.V. (2011). Comparison of environmental risks of pesticides between tropical and nontropical regions. *Integr. Environ. Assess. Manage.*, 7 : 577–586.
- Scelza, R.; Rao, M.A. and Gianfreda, L. (2008). Response of an agricultural soil to pentachlorophenol (PCP) contamination and the addition of compost or dissolved organic matter. *Soil Biol. Biochem.*, 40 : 2162–2169.
- Scopel, E.; Triomphe, B.; Affholder, F.; Da Silva, F.A.M.; Corbeels, M.; Xavier, J.H.V.; Lahmar, R.; Recous, S.; Bernoux, M.; Blanchart, E.; Mendes, D.C.I. and DeTourdonnet, S. (2013). Conservation agriculture cropping systems in temperate and tropical conditions, performances and impacts. A review. *Agron. Sustainable Dev.*, 33 : 113–130.
- Sebiomo, A.; Ogundero, V.W. and Bankole, S.A. (2011). Effect of four herbicides on microbial population, soil organic matter and dehydrogenase activity. *Af. J. Biotech.*, 10: 770–778.
- Shan, M.; Fang, H.; Wang, X.; Feng, B.; Chu, X.Q. and Yu, Y.L. (2006). Effect of chlorpyrifos on soil microbial populations and enzyme activities. *J. Environ. Sc.*, 18: 4–5.
- Sluijsmans, C.M.J. and Kolenbrander, G.J. (1977). The significance of animal manure as a source of nitrogen in soils In: *Proceedings of an International Seminar on Soil, Environment and Fertility Management in Intensive Agriculture*. The Society of the Science of Soil and Manure, Tokyo, 403–441.
- Singh, A.; Singh, M.K. and Ghoshal, N. (2016). Microbial biomass dynamics in a tropical agroecosystem: influence of herbicide and soil amendments. *Pedosphere.*, 26: 257–264.
- Singh, P.; Ghoshal, N. and Singh, R.P. (2011). Influence of Herbicide and Soil Amendments on soil N dynamics, microbial biomass and crop yield in rice–barley sequence under Tropical Dryland Agroecosystems. *Soil Sc. Soc. Am. J.*, 76 : 2208–2220.
- Singh, S.; Ghoshal, N. and Singh, K.P. (2007). Variations in soil microbial biomass and crop roots due to differing resource quality inputs in a tropical dryland agroecosystems. *Soil Biol. Biochem.*, 39 : 76–86.
- Singh, J. and Singh, D.K. (2005). Bacterial, azotobacter, actinomycetes, and fungal population in soil after diazinon, imidacloprid, and lindane treatments in groundnut (*Arachis hypogaea* L.) fields. *J. Environ. Sc. Health B.*, 40 : 785–800.
- Smith, J. and Paul, E. (1990). The significance of soil microbial biomass estimations. *Soil Biol. Biochem.* 6 : 357–396.
- Sofa, A.; Scopa, A.; Dumontet, S.; Mazzatura, A. and Pasquale, V. (2012). Toxic effects of four sulphonylureas herbicides on soil microbial biomass. *J. Environ. Sc. Health B.*, 47 : 653–659.
- Subhani, A.; Ayam, M.E.; Huang, C. and Xu, J. (2000). Effect of pesticides (herbicides) on soil microbial biomass: review. *Pakistan J. Biol. Sc.*, 3 : 705–709.
- Vandana, L.J.; Rao, P.C. and Padmaja, G. (2012). Effect of herbicides and nutrient management on soil enzyme activity. *J. Rice Res.*, 5 :1–2.
- Van Eerd, L.L.; Hoagland, R.E.; Zablutowicz, R.M. and Hall, J.C. (2003). Pesticide metabolism in plants and microorganisms. *Weed Sc.*, 51 : 472–495.
- Vieira, E.F.S.; Cestari, A.R.; Chagas, R.A. and Cortes, G.K.R. (2014). Collection and characterization of suitable matrix for long-release systems—release studies of Atrazine and Diuron herbicides. *Quim. Nova.*, 37 : 398–403
- Vischetti, C.; Perucci, P. and Scarponi, L. (2000). Relationship between rimsulfuron degradation and microbial biomass content in a clay loam soil. *Biol. Fertil. Soils*, 31 : 310–314.
- Vischetti, C.; Casaucci, C. and Perucci, P. (2002). Relationship between changes of soil microbial biomass content and imazamox and benfluralin degradation. *Biol. Fertil. Soils*, 35 : 13–17.
- Vischetti, C.; Coppola, L.; Monaci, E.; Cardinali, A. and Castillo, M.D. (2007). Microbial impact of the pesticide chlorpyrifos on Swedish and Italian biobeds. *Agron. Sustain. Dev.*, 27 : 267–272.
- Volchko, Y.; Norrman, J.; Rosen, L.; Bergknut, M.; Josefsson, S.; Soderqvist, T.; Norberg, T.; Wiberg, K. and Tysklind, M. (2014). Using soil function evaluation in multi-criteria decision analysis for sustainability appraisal of remediation alternatives. *Sc. Tot. Environ.*, 485 : 785–791.

- Wainwright, M. (1978). A review of the effects of pesticides on microbial activity in soil. *J. Soil Sc.*, 29 : 287–298.
- Xia, X.; Zhao, M.; Wang, H. and Ma, H. (2012). Influence of butachlor on soil enzymes and microbial growth. *J. Food Agric. Environ.*, 9 : 753–756.
- Xie, X.M.; Liao, M.; Huang, C.Y.; Liu, W.P. and Abid, S. (2004). Effects of pesticides on soil biochemical characteristics of a paddy soil. *J. Environ. Sc.*, 16 : 252–255.
- Xu, J.; Zhang, Y.; Dong, F.; Liu, X.; Wu, X. and Zheng, Y. (2014). Effects of repeated applications of Chlorimuron-Ethyl on the soil microbial biomass, activity and microbial community in the Greenhouse. *Bull. Environ. Contam. Toxic.*, 92 : 175–182.
- Yang, Y.; Wang, H.; Tang, J. and Chen, X. (2007). Effects of weed management practices on orchard soil biological and fertility properties in southeastern China. *Soil Till. Res.*, 93 : 179–185.
- Zain, N.M.M.; Mohamad, R.B.; Sijam, K.; Morshed, M.M. and Awang, Y. (2013). Effects of selected herbicides on soil microbial populations in oil palm plantation of Malaysia: a microcosm experiment. *Af. J. Microbiol Res.*, 7 : 367–374.
- Zabaloy, M.C.; Garland, J.L.; Go´mez, M.C.; Garland, J.L. and Go´mez, M.A. (2008). An integrated approach to evaluate the impacts of the herbicides glyphosate, 2, 4-D and metsulfuron-methyl on soil microbial communities in the Pampas region, Argentina. *App. Soil Ecol.*, 40 : 1–12.
- Zhao, J.; Ni, T.; Li, Y.; Xiong, W.; Ran, W.; Shen, B.; Shen, Q. and Zhang, R. (2014a). Responses of bacterial communities in arable soils in a rice–wheat cropping system to different fertilizer regimes and sampling times. *PLoS One*. 9 (1) : e85301
- Zhao, J.; Zhang, R.; Xue, C.; Xun, W.; Sun, L.; Xu, Y. and Shen, Q. (2014b). Pyrosequencing reveals contrasting soil bacterial diversity and community structure of two main winter wheat cropping systems in China. *Microb. Ecol.*, 67 : 443–453.