



STUDY OF PHYLOGENY GROUPS OF *ESCHERICHIA COLI* BACTERIA ISOLATED FROM WOMEN VAGINA

Shahad Saad Mohammad, Mohammed Sabri Abdul-Razzaq and Milal Muhammad AL-Jeborry

College of Medicine, Babylon University, Babylon, Iraq.

Abstract

Detection of phylogeny group of *Escherichia coli* isolated from vagina of women suffered from vaginitis infection. In this study, collection about 130 vaginal swabs were obtained from women suffering from vaginitis infection with age (18-40 years). Samples swab were cultured on culture media such as MacConkey and EMB agar for diagnosis, also used molecular method for identification phylogeny groups. Molecular identification based on PCR products were detected by electrophoresis by using 1% agarose for 45 minutes with 70 volt and by using 100bp marker ladder for determine size of amplification PCR products. In this study from 130 swabs isolated only 20(14.8%) isolates diagnosis *E.coli* bacteria after culture on diagnosis culture media while by depending on molecular detection appear 14 (70%) isolates belonging to groups B₂ *Escherichia coli*, 5(25%) isolates belong to group A and 1(5%) belong to group B₁ bacterial type while no appear any isolates belong to group D bacterial type.

Key words : Vaginitis *E. coli* infection, phylogeny, women vagina.

Introduction

Escherichia coli bacteria present in several sites of the body such as vagina and colon and other, *E. coli* that present in vagina has been poorly characterized when compartment with other sites colonization it. These bacteria have ability to causes several infections either symptomatic or asymptomatic, also studies appear present relationships between *Escherichia coli* normal flora in genital tract and those causing intra-amniotic infection (Sáez-López *et al.*, 2016).

There are four phylogeny *E. coli* groups (A, B₁, B₂, D) (Carlos *et al.*, 2010; Clermont *et al.*, 2013). The *chuA* is gene responsible for iron transport in enterohemorrhagic O157 : H7 while *yjaA* gene play essential role in cellular response to hydrogen peroxide and acid stress and the function of *TspE4.C2* is not yet understand (Lee, 2011).

However, *Escherichia coli* strains categorized in four phylogeny groups, the strain that causes extra-intestinal infection are acute strain that belong to group B₂ and D, while symbiotic group are belong to A₁ and B₁ (Clermont and Bonacorsi, 2011; Baponi *et al.*, 2016).

Nagarjuna *et al.* (2015) show the commensal *E. coli* strains belong to groups A and B₁, whereas the strain

causes extra-intestinal infection belong to group B₂ and D. The intestinal *Escherichia coli* are mixture of all phylogenetic groups and may consider as a reservoir for the pathogenic isolates.

Materials and Methods

Patients

130 vaginal swabs collected from women suffering from vaginal and urinary tract infections with age (18-40 years) were submitted to hospital of maternity and children in Hilla city through period from (October 2016 to April 2017).

Laboratory diagnosis and identification of bacterial isolate

Vaginal swabs collection immediately inoculation into transport media tube for preserved from dryness until transport to laboratory, then the swabs were culture on growth media such as MacConkey and EMB agar and incubation at 37°C for 24 hours. By depended on (MacFaddin, 2000) diagnosis procedure recommended take single colony from each positive culture and noting the morphology properties (color production, colony shape texture and edge). Also using Gram stain procedure for

detection bacterial belong to Gram positive or Gram negative (Winn *et al.*, 2006).

Molecular identification

DNA was extracted by using gene aid kit specific for DNA extraction from Gram negative bacteria as *E. coli* belong to gram negative bacterial type and accordance with gene aid protocol.

Detection of phylogeny groups by PCR

The major phylogenetic groups of *E. coli* (A, B₁, B₂ and D) were identification by depended on specific genes *chuA*, *yjaA* and DNA fragment *TspE4.C2* (Clermout *et al.*, 2000).

The primer used were *chuA*, *TspE4.C2* and *yjaA* which generated 279, 152 and 211 bp fragment, respectively. The information of the three PCR amplification result in assignment of the isolates to phylogenetic groups as follows : *chuA*⁺, *yjaA*⁺ group B₂; *chuA*⁺, *yhaA*-group D; *chuA*⁻, *TspE4.C2*⁺, group B₁ and *chuA*⁻, *yjaA*⁺, *TspE4.C2*⁻, group A. The polymerase chain reaction products size were determined by using molecular marker 100pb.

Results

Twenty (14.8%) vaginal *E. coli* (VEC) isolates were recovered from 135 female suffering from vaginitis and urinary tract infection. All vaginal *E. coli* isolates were subjected to phylogrouping by PCR.

In this study, it was noticed that *E. coli* isolated from women with vaginitis belongs mainly to the phylogeny group B₂ (14/20) followed by the phylogeny group A (5/20) and only one isolate (1/20) for group B₁.

Detection of *ChuA*

All *E. coli* 20 (14.8%) isolates are subjected for detection of gene *chuA* by using specific primer at 279 bp. It was found that 14 isolates gave positive for this marker as shown in fig. 1.

Detection of *yjaA*

All *E. coli* isolates are subjected for detection of *yjaA* gene by using specific primer at 211 bp. It was found that 17 isolates gave positive for this Marker as shown in fig. 2.

Detection of *TspE4.C2*

All *E. coli* isolates are subjected for detection of *TspE4.C2* gene by using specific primer at 152 bp. It was found that 14 isolates gave positive for this marker as shown in fig. 3.

Concisely, the results in this table showed that most *E. coli* isolate, belong to group B₂ 14 isolate (70%)

Table 1 : Contents of the reaction mixture.

No.	Contents of reaction mixture	Volume
1)	green master mix	12.5 µl
2)	forward primer	2.5 µl
3)	reverse primer	2.5 µl
4)	DNA template	5 µl
5)	Nuclease free water	2.5 µl
Total volume		25 µl

Table 2 : Distribution of *E. coli* isolated according to phylogeny groups.

Phylogeny groups	<i>TspE4.C2</i>	<i>yjaA</i>	<i>chuA</i>	No. of isolates
A	-	-	-	1
B ₂	+	+	+	2
B ₂	+	+	+	3
B ₂	+	+	+	4
B ₂	+	+	+	5
B ₂	+	+	+	6
B ₁	+	+	-	7
B ₂	+	+	+	8
B ₂	+	+	+	9
B ₂	+	+	+	10
A	-	-	-	11
B ₂	+	+	+	12
B ₂	+	+	+	13
B ₂	+	+	+	14
B ₂	-	+	+	15
A	-	+	-	16
A	-	-	-	17
B ₂	+	+	+	18
A	-	+	-	19
B ₂	+	+	+	20

followed by group A (5 isolates) (25%) and only one for B₁ only one isolate (5%).

Discussion

The presence of *E. coli* in healthy vagina and also in patients with vaginitis may have many explanation where the source of its existence in vagina may be as a contaminant from faces or it may be true pathogen of this disease, as a result of opportunistic capability of this bacteria to do various type of infections in women such as UTI and vaginitis (Ahmed, 2015).

The phylogenetic groups of *E. coli* isolated from vagina was detected by identifying the presence of specific PCR amplified fragment (*chuA*, *yjaA* and *TspE4.C2*).

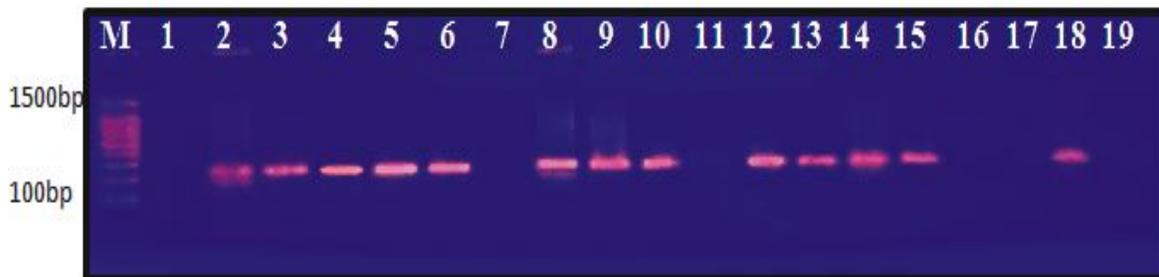


Fig. 1 : Gel electrophoresis of PCR product of *chuA* gene (1-19) : No. of isolates obtained from vaginal samples. M= Molecular marker (100bp).

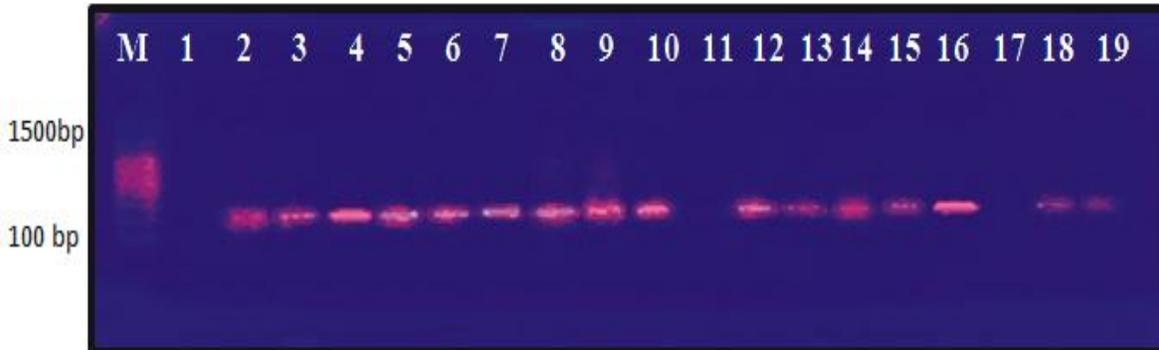


Fig. 2 : Gel electrophoresis of PCR product of *yjaA* gene(1-19) : no. of isolates obtained from vaginal samples . M= Molecular marker (100bp).



Fig. 3 : Gel electrophoresis of PCR product of *TspE4.C2* gene(1-19) : no. of isolates obtained from vaginal samples. M= Molecular marker (100bp).

All the fourteen isolates belonging to group B₂ are considered as extra-intestinal and pathogenic to man, and this will confirm their role in vaginitis. Whereas the isolates belong to group A and B₁ are considered as commensal, not true pathogen that they may be isolated as a contaminants from anal area. However, no isolates was classified within the group D.

The study carried out by Abdul-Lateef (2011), who found that most *E. coli* isolates belong to the group B₂ (30%), A (30%) & D (30%). At the same rate this is not identical to the results of this study which no isolates was found to be related to group D. According to the data obtained. *E. coli* isolates were distributed in four phylogeny groups depending on the results of the presence and absence of phylogeny genes (*chuA*, *yjaA*,

TspE4.C2).

Group D is not found in this study, although many studies confirmed that most *E. coli* strains that are isolated from extraintestinal regions are either belong to phylogeny group B₂ or D (Da Silva and Mendonca, 2012; Rasmussen *et al.*, 2012; Johnson *et al.*, 2012). But this study showed that most isolates of vaginal *E. coli* belong to the group B₂. However, Al Safar (2016) have found all vaginal *E. coli* belong to the group B₂ not for A, B₁ or D. However, other studies (Al-Khaqani *et al.*, 2017; Guiral *et al.*, 2011; Usein *et al.*, 2011) have found that most vaginal *E. coli* belong mainly to the group B₂ that may indicate that source of these isolates is not from the faeces.

These results may highlight the importance of *E. coli* as a pathogen in the Vaginitis with high risks of transmission to the upper parts of the genital tract and, if the women are pregnant, it may cause serious infections to the fetus, or the newborn (Al-Khalide *et al.*, 2015).

The results of this study appeared that the *chuA* gene was present in all isolates belonging to groups B₂ and D and was absent from all isolates belonging to groups A and B₁. The *yjaA* gene allowed perfect distinguish between group B₂ and group D and it was present in all isolates belonging to group A. Also, the *TspE4.C2* is present in group B₁ strains and absent from all group A isolates (Abdul-Razzaq and Abdul-Lateef, 2011).

Phylogeny groups was detect the source of infection where the groups A& B₁ are commensal, so the source of *E. coli* in cases of vaginitis may be the intestinal because *E. coli* is commensal mostly in human intestinal (Cribby *et al.*, 2008).

Conclusion

Most bacterial isolates belong to phylogeny group B₂ and then to group A.

Recommendation : Study the phylogeny groups and its relation to gene conversion in bacteria.

References

- Abdul-Razzaq, M. S. and L. A. Abdul-Lateef (2011). Molecular phylogeny of *Escherichia coli* isolated from clinical samples in Hilla, Iraq. *Afr. J. Biotechnol.*, **10(70)** : 15783-15787.
- Ahmed, M. M. (2015). Comparative Study of Molecular Phylogeny, Adhesion Genes and Antiobiogram of *Escherichia Coli* Clinical Isolates From High Vaginal Swabs and Urine in Women. *Karbala J. Med.*, **8(1)**.
- Abdul-Lateef, L. A. (2011). Pylogeny and Pathogenicity Islands among *Escherichia coli* Isolated from clinical cases. *Ph.D. Thesis*. College of Medicine, University of Babylon. Iraq.
- Al-Khalide, E. K., M. M. Ahmed and Z. H. Abood (2015). Virulence factors genes and phylogenic groups of *Escherichia coli* isolated from high vagina and endocervix of women from Kerbala. *Karbala Journal of Medicine*, **8** : 2292-2296.
- Al-Saffar, A. K. H. (2016). Determination of *Escherichia coli* phylogenetic group isolated from women with vaginitis in Hilla City, Iraq. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, **7** : 1467-1470.
- Al-Khaqani, M. M., M. S. Alwash and H. O. Al-Dahmoshi (2017). Investigation of phylogroups and some virulence traits among cervico-vaginal *Escherichia coli* (CVEC) isolated for female in Hilla City, Iraq. *Malays. J. Microbiol.*, **13(2)** : 132-138.
- Baponi, S., A. Taravati and M. Dilmagani (2016). Determination of phylogenetic groups of *Escherichia coli* isolated from human urine in Urmia city. *Crescent Journal of Medical and Biological Sciences*, **3(3)** : 97-99 ref.13.
- Clermont, O., S. Bonacorsi and E. Bingen (2000). Rapid and simple determination of *Escherichia coli* phylogenetic group. *Appl. Environ. Microbiol.*, **66** : 4555-4558.
- Carlos, C., M. M. Pires, N. C. Stoppe, E. M. Hachich, M. I. Z. Sato, T. A. T. Gomes, L. A. Amaral and L. M. M. Ottoboni (2010). *Escherichia coli* phylogenetic Group Determination and its Application in the Identification of the Major Animal Source of Fecal Contamination. *BMC Microbiology*, **10** : 161.
- Clermont, O., J. K. Christenson, E. Denamur and D. M. Gordon (2013). The Clermont *Escherichia coli* phylotyping method revisited: improvement of specificity and detection of new phylo-groups. *Environmental Microbiology Reports*, **5** : 58-65.
- Clermont, O. and S. Bonacorsi (2011). Bingen ERapid and simple determination of the *Escherichia coli* phylogenetic group. *Appl Environ Microbiol.*, **200(66)** : 4555-4558.
- Cribby, S., M. Taylor and M. Reid (2008). Vaginal Microbiota and the Use of Probiotics. *Interdisciplinary Perspectives on Infectious Diseases*. Article ID 256490, 9 pages.
- Da Silva, G. J. and N. Mendonca (2012). Association between antimicrobial resistance and virulence in *Escherichia coli*. *Virulence*, **3(1)** : 18-28.
- Guiral, E., J. Bosch, J. Vila and S. M. Soto (2011). Prevalence of *Escherichia coli* among samples collected from the genital tract in pregnant and nonpregnant women: relationship with virulence. *FEMS Microbiol Lett.*, **314** : 170-173.
- Johnson, J. R., C. Urban, S. J. Weissman and J. H. Jorgensen (2012). Molecular epidemiological analysis of *Escherichia coli* sequence Type ST131 (O25:H4) and blaCTX-M-15 among extended spectrum- β -lactamase-producing *E. coli* from the United States, 2000 to 2009. *Antimicrob. Agents Chemother.*, **56(5)** : 2364-2370.
- Lee, C. C. Y. (2011). Genotyping *Escherichia coli* Isolates from Duck, Goose and Gull Fecal Samples with Phylogenetic Markers using Multiplex Polymerase Chain Reaction for Application in Microbial Source Tracking. *Journal of Experimental Microbiology and Immunology*, **15** : 130 - 135.
- MacFaddin, J. F. (2000). Biochemical tests for identification of medical bacteria. 3rd ed. Williams and Wilkins-Baltimore, pp: 321-400.
- Nagarjuna, D., G. Mittal, R. S. Dhanda, P. K. Verma, R. Gaiind and M. Yadav (2015). Faecal *Escherichia coli* isolates show potential to cause endogenous infection in patients admitted to the ICU in a tertiary care hospital. *PMC4522595*, **4(7)** : 57-66.
- Rasmussen, L. S., K. Ejrnaes, B. Lundgren, A. M. Hammerum and N. F. Moller (2012). Virulence factors and phylogenetic

- grouping of *Escherichia coli* isolates from patients with bacteremia of urinary tract origin relate to sex and hospital- vs. community-acquired origin. *Int J Med Microbiol.*, **302**: 129-134.
- Sáez-López, E., A. Cossa, R. Benmessaoud, L. Madrid, C. Moraleda, S. Villanueva, H. Tligui, B. Moiane awnd H. Alami (2016). Characterization of Vaginal *Escherichia coli* Isolated from Pregnant Women in Two Different African Sites. *Marcia Edilaine Lopes Consolaro*, **11(7)**: e0158695.
- Usein, C. R., L. A. Grigore, R. M. Georgescu, M. C. Băltoiu, M. Condei and M. D. Telean (2011). Phylogenetic background and extraintestinal virulence genotypes of *Escherichia coli* vaginal strains isolated from adult women. Davila, Bucharest, Romania. *Revista Română de Medicină de Laborator*. **19(¼)**.
- Winn, W. C., S. D. Allen, W. M. Janda, E. W. Koneman, G. W. Procop, P. C. Schreckenberger and G. L. Woods (2006). *Koneman's color atlas and textbook of diagnostic microbiology*. 6th ED. Lippincott Williams and Wilkins, USA. PP:234-241.